

FATE OF NITROUS OXIDE (N₂O) IN FINNISH BOREAL FOREST AND PEATLAND SOILS

Sankari Rajasakaren

Master of Science Thesis

MSc degree in Environmental Biology

University of Eastern Finland (Kuopio), Faculty of Science and Forestry

May 2017

Abstract

Recent studies show that nitrous oxide (N_2O) has a missing sink in soils but the potential of this sink of N_2O is poorly understood. Denitrification has been shown to be the major consumption process of N_2O and could result as a sink potential for N_2O if the process is highly efficient. However, the process of denitrification is influenced by several factors such as oxygen, pH, temperature, nutrient availability, soil moisture content and soil type. The main aim of this thesis was to understand the mechanisms that influence N_2O concentrations in boreal forest soils and peatlands and to evaluate their potential to act as sinks for N_2O . The boreal forest study site selected for this study is located in Heinävesi and the peatland site in Salmisuo. Soil samples from the organic and mineral layer of the boreal forest and the upper 0-10cm and lower 10-20cm of the peatlands were collected to understand the differences in N_2O concentrations within different soil layers. Furthermore, the factors affecting N_2O concentrations were investigated, namely; the role of oxygen and substrate availability (more specifically, NH_4^+ , NO_3^-). The effect of different carbon substrates namely succinate, butyrate, formate, propionate and lactate on N_2O concentrations were also investigated to understand if they stimulated distinct microorganisms which influenced net N_2O consumption/production potential.

In anoxic treatment both boreal peat (Salmisuo) and forest (Heinävesi) soil showed a net consumption of N_2O . However, there were differences in net N_2O consumption potential between forest soils and peat as well as between the different layers within each site. In oxic treatment forest soils showed a net production of N_2O in organic soil but near negligible production of N_2O in mineral soils. However in peat, there was a net consumption of N_2O in both peat layers. The net consumption potential of N_2O in peat was smaller in oxic conditions than in anoxic conditions. The addition of different carbon substrates to the mineral (Heinävesi) forest soil and the lower 10-20cm peat (Salmisuo) soil resulted in similar trends with exception of formate. Although there were similar trends observed in both soils, the consumption or production potential depending on the relevant carbon substrate was comparatively larger in the lower 10-20cm (Salmisuo) peat.

This study indicates that the effect of oxygen on denitrification plays a crucial role in regulating N_2O fluxes in boreal soils and their potential to act as sinks for N_2O . These results

also indicate that boreal soils are sensitive to aerobic/anaerobic conditions which can influence their potential to act as a sink for N₂O.

Acknowledgements

This master's thesis represents to me a significant milestone in my life. It has been a period of intense learning, not only in the scientific aspects but also on a personal level. My time at University of Eastern Finland (UEF) at Kuopio has been nothing short of amazing. It has been the place that has provided me with unique opportunities since I came as an undergraduate exchange student from Germany in the autumn of 2011. Throughout my years at UEF, I have met many remarkable people who have been invaluable in my academic journey.

I would first like to express my sincere appreciation and wholehearted gratitude to my principle supervisor Jenie Gil Lugo. Her invaluable guidance and moral support helped me to complete this master thesis successfully. I am very much obliged to her for listening to all my concerns and even providing me with emotional support when I went through rough patches in completing this thesis. Ms Lugo always answered all my queries and made me feel comfortable in exploring my subject matter in the manner I chose to. She was also an important source of inspiration for me in completing this thesis.

I would also like to thank my second supervisor Professor Toini Holopainen for her invaluable guidance. I am extremely lucky and grateful that I was able to attain her in the reading of this manuscript. Professor Holopainen has been a tremendous beacon of light for me during this journey and without her support I would not have been able to complete a bachelor or master's thesis. Albert Einstein once said, 'It is the supreme art of the teacher to awaken joy in creative expression and knowledge'. This quote succinctly and aptly explains the high esteem I have for Professor Holopainen.

I would also like to thank my primary 6 school teacher, Mrs Chu-Tan Kim Yang. She made it fundamentally possible for me to enter the best schools in Singapore which enabled me to enter university. She encouraged her students to dream big and dream the impossible. I will forever be grateful to her; for I have always applied all the lessons she has taught me in life. I also owe much of my excellence to her.

Finally, I would like to thank my family and friends. My parents have provided me with limitless support and confidence in me which I cherish so dearly. They took the plunge with me in this journey since I arrived in Europe as a young, naive 19 year old. Nothing could have been possible without their love and encouragement. I am aware every single day how lucky I am to have parents like them who have constantly supported my dreams. Since I was a child,

my brother Balamuralli has been my role model. He is another reason why I was able to study abroad and gave me ample confidence to overcome any obstacle. I could not have asked for a better person to inspire me throughout my life. I would also like to thank my fiancé, Pierre-Emmanuel Panouillot. He has been extremely supportive and helpful throughout this entire journey and I am lucky to have met him in Kuopio. His unyielding friendship, love and support have enabled me to succeed. I cherish the numerous moments where he cared for me and helped me in preparing this thesis. He has always been there in all my victories and defeats and I could not have asked for a better partner in life. I would also like to thank all my friends and colleagues at NorskHydro in Norway who have been incredible. They are not simply work colleagues, superiors or friends but they are family. Their support has made this thesis possible. I owe my eternal gratitude for everything they have done for me.

Dedicated to my parents, Mr and Mrs Rajasakaren who are the most supportive and loving parents.

Contents

| | |
|---|-----------|
| 1. Introduction | 8 |
| 1.1 Nitrous oxide | 8 |
| 1.2 Research Objectives and Hypotheses | 9 |
| | |
| 2. Literature Review | 10 |
| 2.1 Boreal Region..... | 10 |
| 2.2 Processes that result in production and consumption of N ₂ O in soils..... | 12 |
| 2.3 Atmospheric N ₂ O fluxes from boreal soils | 14 |
| 2.4 Factors affecting N ₂ O sinks..... | 16 |
| 2.4.1 Effects of soil oxygen on N ₂ O fluxes..... | 16 |
| 2.4.2 Effects of soil pH on N ₂ O fluxes..... | 16 |
| 2.4.3 Effect of soil temperature on N ₂ O fluxes in natural ecosystems..... | 17 |
| 2.4.4 Effect of nutrient availability on N ₂ O fluxes | 17 |
| 2.4.5 Effects of soil moisture content..... | 18 |
| 2.4.6 Effect of soil type on N ₂ O fluxes | 19 |
| 2.4.6.1 Soil Carbon-to-Nitrogen ratio (C:N ratio)..... | 19 |
| 2.4.6.2 N ₂ O emissions from Organic vs mineral soil types | 20 |
| 2.4.6.3 Natural vs altered ecosystems | 20 |
| 2.4.7 Hole in the pipe model | 21 |
| | |
| 3. Materials and Methods | 22 |
| 3.1 Study site | 22 |
| 3.2 Experimental set-up..... | 23 |
| 3.2.1 Effect of oxygen on N ₂ O, NH ₄ ⁺ and NO ₃ ⁻ concentrations | 23 |
| 3.2.2 Effect of electron donors on mineral forest soil and lower 10-20cm peat soil on N ₂ O concentrations in oxic conditions..... | 25 |
| 3.3 Statistical Analysis | 25 |
| | |
| 4. Results | 26 |
| 4.1 Soil properties of the study sites | 26 |
| 4.2 Effects of Anoxic conditions on N ₂ O fluxes in forest soils (Heinävesi)..... | 27 |

| | |
|---|-----------|
| 4.3 Effects of Oxic conditions on N ₂ O fluxes in forest soils (Heinävesi)..... | 33 |
| 4.4 Effects of anoxic conditions on N ₂ O fluxes in peat (Salmisuo)..... | 35 |
| 4.5 Effects of Oxic conditions on N ₂ O concentrations in peat (Salmisuo)..... | 37 |
| 4.6 Effects of different electron donors on N ₂ O fluxes from the mineral layer of Heinävesi forest and the lower peat layer (10-20cm) of Salmisuo | 39 |
| | |
| 5. Discussion | 40 |
| 5.1 Effects of anoxic conditions on N ₂ O fluxes in forest soils (Heinävesi)..... | 40 |
| 5.2 Effects of oxic conditions on N ₂ O fluxes in forest soils (Heinävesi)..... | 41 |
| 5.3 Effects of Anoxic conditions on N ₂ O fluxes in peat (Salmisuo)..... | 42 |
| 5.4 Effects of Oxic conditions on N ₂ O fluxes in peat (Salmisuo)..... | 43 |
| 5.5 Effect of different electron donors on net N ₂ O consumption/potential rate in oxic conditions in mineral forest soil and peat..... | 44 |
| | |
| 6. Conclusion | 46 |
| | |
| 7. References | 47 |

1. Introduction

1.1 Nitrous oxide

Nitrous oxide (N_2O) is a potent greenhouse gas (GHG) contributing to climate change with an atmospheric lifetime of 114 years (Forster *et al.*, 2007). With its ozone-depleting properties (Ravishankara *et al.*, 2009) and a global warming potential 300 times that of carbon dioxide (CO_2), N_2O is attributed to be the third most important contributor to radiative forcing (Forster *et al.*, 2007). As N_2O plays an important role in the atmosphere, identifying the sources and sinks of this greenhouse gas is critical for understanding its implications across ecosystems. Global N_2O emissions are estimated to be $17.71 \text{ Tg N yr}^{-1}$ (Schlesinger, 2013). The main sources of N_2O to the atmosphere are agricultural (2.8 Tg N yr^{-1}) and tropical rain forest soils (4.0 Tg N yr^{-1}) contributing around 40% of the total emissions. Since the pre-industrial era, global atmospheric N_2O concentrations have increased by about 20%, which has been attributed largely to the increased use of nitrogen-based fertilizers (IPCC, 2013). It has also been predicted that with continuous increase of N_2O in the atmosphere, the significance of N_2O to greenhouse effect and ozone depletion will be more severe in the twenty-first century (Ravishankara *et al.*, 2009).

The most important and main sink for N_2O is the photochemical reactions ($12.5 \text{ Tg N yr}^{-1}$), which take place in the stratosphere (Crutzen and Oppenheimer, 2008). However, there is growing evidence that soils can also behave as sinks for N_2O (Chapuis-Lardy *et al.*, 2007; Schlesinger, 2013). Nevertheless, further understanding is necessary to elucidate the potential of soils as a sink of N_2O . Soils prone to high N_2O sink activity are mostly located in northern regions (Kroeze *et al.*, 2007). Although Schlesinger (2013) has estimated global N_2O uptake in soils to be not higher than 0.3 Tg N yr^{-1} which corresponds to global sinks being no greater than 2% of estimated sources of atmospheric N_2O , the exact potential of soils as N_2O sinks have yet to be exactly determined. Hence, by understanding the mechanisms that influence N_2O uptake in soils we can ascertain the atmospheric importance of this N_2O sink.

In a review investigating 100 different studies on natural or recovering ecosystems, measured uptake potentials of N_2O in soils ranged from $1.0 \mu\text{g Nm}^{-2} \text{ h}^{-1}$ to $207 \mu\text{g Nm}^{-2} \text{ h}^{-1}$ with a median value of $4 \mu\text{g Nm}^{-2} \text{ h}^{-1}$ with the highest consumption of N_2O observed in wetland and peatland ecosystems (Schlesinger, 2013). The net fluxes of N_2O indicate whether soils are a net source or sink with N_2O production and consumption primarily related to the processes of nitrification and denitrification, respectively. These in turn depend on environmental factors

regulating consumption and production. Recent studies show that N₂O has a missing sink in soils but the potential of sinks of N₂O have been poorly delineated. It is pertinent to note that denitrification process can result as a sink potential for N₂O if the process is efficient.

For sink activity to occur, the ambient N₂O concentration in the atmosphere has to exceed that in the soil gas phase. The N₂O exchange between soils and the atmosphere can be explained by the equilibrium concept (Conrad and Dentener, 1999). At N₂O compensation point, the concentration of N₂O in the soil gas phase results in no net exchange of N₂O between ambient atmosphere and soil gas phase. Hence, it can be concluded that the compensation point determines the direction of the flux at a given ambient atmospheric N₂O concentration. Compensation concentrations of N₂O are typically significantly higher than the ambient atmospheric concentrations which correlate with the observation that most soils are sources of atmospheric N₂O. Studies that indicate consumption of N₂O by soils suggest that during uptake the compensation concentrations are lower than ambient concentrations, i.e. <310ppbv (Kroeze *et al.*, 2007). It has also been suggested that the compensation concentration for N₂O increases with other factors such as temperature, soil moisture and nitrogen availability (Conrad and Dentener, 1999).

1.2 Research Objectives and Hypotheses

In this study, the main objective was to quantify the N₂O fluxes at a boreal spruce forest and pristine peatland in Finland in anaerobic and aerobic conditions. Two main hypotheses were tested. The first was that aerobic conditions in forest soils would result in markedly increased N₂O fluxes in organic soils compared to mineral soils; but in both peat layers would result in consistent N₂O fluxes. The second hypothesis was that the addition of different carbon sources to both forest soils and peat would affect N₂O fluxes because of the resulting preference for certain carbon sources by microbes in the soils. To test these hypotheses experimental anaerobic and aerobic conditions and manipulation of carbon sources were arranged in flask microcosms and following N₂O fluxes were determined. Furthermore, mineral nitrogen (nitrate and ammonium) concentrations in soil were quantified.

2. Literature Review

2.1 Boreal Region

The boreal region is the biggest terrestrial biome in the world forming a distinct band of vegetation. It comprises nearly 14.5% of the Earth's terrestrial ecosystems (Berner *et al.*, 2015). The boreal region (Figure 1) is a circumpolar range throughout the entire Northern Hemisphere from Alaska and Canada; across the Atlantic to Norway, Sweden and Finland, through vast areas of Russia, and into parts of China, Mongolia, the Korean Peninsula and northern Japan (Miyawaki and Tüxen, 1977). It is characterized by harsh winters with mean temperatures below freezing for up to six months where much of boreal life comes to a halt (Runesson, 2011). Meanwhile, summers in boreal forests are characterized by long, warm periods of sunlight which leads to swift growth in boreal forests (Rumney, 1968 and Larsen, 1980). In Finland, the biogeography is composed of extensive boreal forests, mires and peatlands.

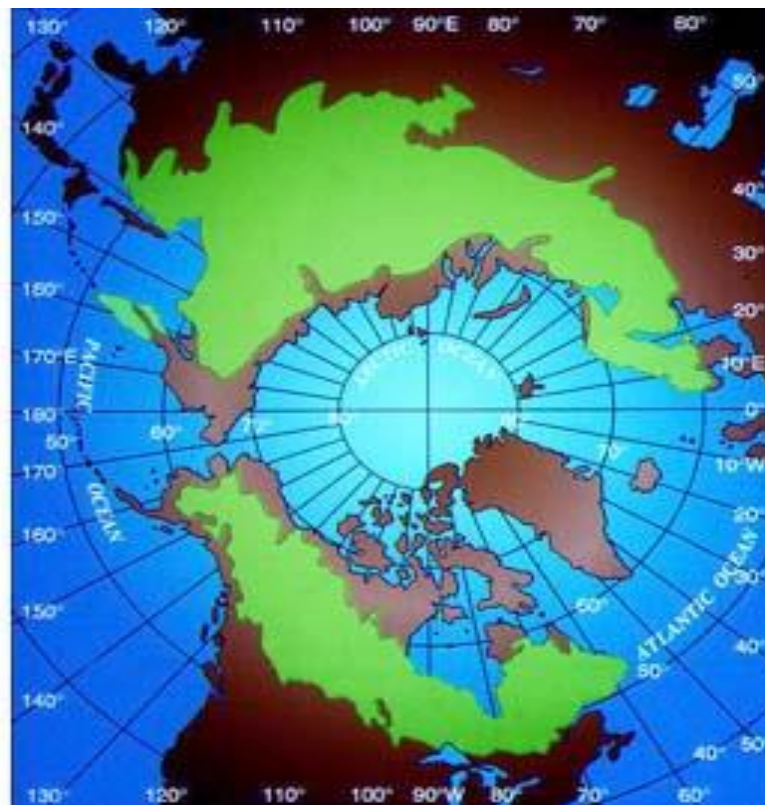


Figure 1: Circumpolar Range of Boreal Region (**indicated in green**) (Hare and Ritchie, 1972)

The boreal forests that dominate Finland are characterized by *Pinus Silvestris* (Pine), *Picea abies* (Spruce) and *Betula pendula* and *B. Pubescens* (Birch) which together encompass 73% of Finnish land surface (METLA, 2013). The boreal forests are dominated by nutrient-poor

soils, with slow nutrient cycling. This is because of limited availability of nitrogen due to low mineralization rate and the high demand for available nitrogen for growth (Näsholm *et al.*, 1998). Although the growth of boreal forests is generally N-limited, wide ranges in nutrient availability and interaction between elements have been observed (Vitousek *et al.*, 1997). Hence, it is important to understand nitrogen cycling processes thoroughly.

Peatlands are defined as a subset of wetlands containing peat-covered terrain having a minimum depth of peat of 30cm (Joosten and Clarke, 2002). Peat is a soil type, consisting of partially decomposed organic matter accumulating under water-saturated conditions due to incomplete decomposition. Peat forms as a result of anoxic condition, low decomposability of plant material and other factors. This accumulation of peat results in more acidic and nutrient-poor conditions because the influence of cations derived from mineral soil decreases over time (Vitt *et al.*, 2005)

There are two main types of peatlands, minerotrophic and ombrotrophic. In minerotrophic peatlands, also known as fens, water in the peat surface is connected with, or has passed over or through surrounding mineral soil (Rydin and Jeglum, 2013). Fens have slightly acidic to slightly basic mineral-rich waters from groundwater discharge and seepage. They may be flat or gently sloping and are dominated by sedges, grasses, and non-sphagnum mosses. Fens can be classified in poor fen, intermediate-moderately rich fen and extremely rich fen based on their vegetation composition, local geology, chemical and nutrient status of the water supply among others. Poor fens have a pH of 4-5.5, while intermediate-moderately rich fens have a pH of 5-7, and rich fens have a pH of 6.8-8. On the other hand, ombrotrophic peatlands, also known as bogs, are isolated from mineral soil water and only receive their water and nutrients from precipitation. Bogs are nutrient poor and usually have a pH range of 3.5-4.2. They are dominated by the species of moss genus *Sphagnum* and heath shrubs (Rydin and Jeglum, 2013). Fens have a broader range of nutrient and pH variation and therefore more abiotic and biotic variation than bogs (Rydin and Jeglum, 2013).

In Finland, peatlands typically contain large amounts of carbon (C) and nitrogen (N) in organic matter in soil and vegetation (Gorham, 1991). Hence, they play an important role in the global C and N cycles particularly with processes related to producing and consuming greenhouse gases (GHG) (Khalil, 1999).

2.2 Processes that result in production and consumption of N₂O in soils

N₂O emissions from the soil can be largely attributed to bacterial and fungal respiratory processes in soils, namely denitrification and nitrification (Lassey and Harvey, 2007). About 70% of global N₂O emission have been attributed to microbial processes of nitrification and denitrification in soils (Braker and Conrad, 2011; Syakila and Kroeze, 2011). However, it is impossible to solely ascribe the sources of N₂O to microbial nitrification and denitrification because several biogeochemical pathways and abiotic mechanism produce or consume N₂O (Butterbach-Bahl *et al.*, 2013). Overall, soils are a complex heterogeneous matrix where N₂O production can occur through different processes simultaneously in different microsites of the same soil (Butterbach-Bahl *et al.*, 2013).

During the process of nitrification, there is microbial oxidation of ammonium (NH₄⁺) to nitrate (NO₃⁻) by chemoautotrophic nitrifying bacteria in aerobic conditions. This process is affected by numerous factors including NH₄⁺, nitrite (NO₂⁻), phosphate (PO₄³⁻) concentrations, oxygen (O₂), pH, temperature, water potential and possible allelopathic compounds (Haynes, 1975). In aerobic autotrophic nitrification, there is stepwise oxidation of ammonia (NH₃) to hydroxylamine (NH₂OH) and subsequently to NO₂⁻ and NO₃⁻ (Kowalchuk and Stephen, 2001). N₂O production from these processes can result because of enzymatic decompositions of the substrates NH₂OH and NO₂⁻ (Arp and Stein, 2003; Baggs, 2011). Biosynthesis of the intermediate NO₂⁻ as substrate for soil N₂O production can occur during the process of nitrate ammonification and nitrifier denitrification (Poth and Focht, 1985; Baggs, 2011; Wrage *et al.*, 2001). Chemodenitrification also contributes to N₂O production when nitrite ion is chemically reduced to N₂O by compounds such as amines present in soil organic matter, and by inorganic ions (Fe²⁺, Cu²⁺), particularly in subsoils (Granli and Bøckman, 1994).

Denitrification is a form of anaerobic respiration whereby the sequential dissimilatory reduction of NO₃⁻ to NO₂⁻, nitric oxide (NO), N₂O and dinitrogen (N₂) is catalyzed by nitrate reductases (NaR or Nap), nitrite reductase (NiR), nitric oxide reductase (NoR) and nitrous oxide reductase (N₂OR) respectively (Zumft, 1997). Denitrification is traditionally considered to require completely anoxic conditions because O₂ is seen as a competing electron acceptor for NO₃⁻ respiration and key enzymes of the denitrification pathways are inhibited by relatively small amounts of O₂ (Zumft, 1997; Shapleigh, 2006). It is dominated by heterotrophic bacteria and fungi with the process considered to be the main N₂O producing process (Skiba and Ball, 2002). It is mainly affected by the presence of bacteria possessing

metabolic capacity; the availability of suitable reductants such as organic carbon; the restriction of O₂ availability and the availability of N oxides such as NO₃⁻, NO₂⁻, NO or N₂O. Closely coupled with denitrification, microbial co-denitrification can utilize N from organic co-substrates for increased N₂O production (Kim *et al.*, 2012).

In recent greenhouse gas models, the final step of denitrification (N₂O → N₂) has been assumed to be the major controlling mechanism reducing N₂O fluxes to the atmosphere (Sanford *et al.*, 2012). Most N₂O is produced when there is incomplete denitrification in suboxic and anoxic conditions whereby anaerobic microbial bacteria reduce nitrate to nitrite and subsequently to gaseous nitrogen compounds nitric oxide (NO) and N₂O. In table 1, adapted from Firestone and Davidson (1989), the factors that affect the proportion of N₂O produced relative to N₂ in denitrifying cells and soils are shown.

Table 1: Factors affecting the proportion of N₂O and N₂ produced during denitrification as adapted from Firestone and Davidson (1989)

| Factor | Will increase N ₂ O/N ₂ |
|--|---|
| [NO ₃ ⁻] or [NO ₂ ⁻] | Increasing oxidant |
| O ₂ | Increasing O ₂ |
| Carbon | Decreasing C availability |
| pH | Decreasing pH |
| [H ₂ S] | Increasing sulphide |
| Temperature | Decreasing temperature |
| Enzyme status | Low N ₂ O reductase activity |

For N₂O to be produced, the availability of oxidant (N-oxide) has to exceed the availability of reductant (organic carbon) resulting in higher proportion of N₂O to N₂ (Table 1). Moreover, decreasing temperature and lower N₂O reductase activity also results in the same phenomenon (Table 1). The ratio of N₂O/NO₃⁻ has been observed to increase with lowering of oxygen partial pressure and increasing acidity (Goreau *et al.*, 1980; Martikainen, 1985).

Although denitrification is traditionally believed to be inhibited by O₂, there is substantial evidence for co-respiration or co-metabolism of O₂ and NO₃⁻ by different genera of cultured bacteria within the *Alpha*-, *Beta*- and *Gammaproteobacteria* under both alternating oxic/anoxic phases and fully aerated conditions (Robertson and Kuenen, 1984; Lloyd, *et al.*,

1987; Bonin and Gilewicz, 1991; Baumann *et al.*, 1996). In bacterial cultures, N₂O and N₂ are produced when O₂ concentrations range from near anoxic to c.90% air saturation (Robertson and Kuenen, 1984; Körner and Zumft, 1989). N₂O produced during nitrification and denitrification contains some differences. N₂O produced during nitrification is more depleted in ¹⁵N and ¹⁸O relative to substrates while N₂O produced during denitrification is less depleted (Butterbach-Bahl *et al.*, 2013).

Nitrogen oxides can be reduced by bacteria, archaea and fungi groups (Philippot *et al.*, 2007; Hayatsu *et al.*, 2008). These micro-organisms can be classified as complete denitrifiers and non-denitrifying N₂O reducers. Complete denitrifiers are facultative aerobes which are ecophysiologicaly homogenous with the potential to switch from oxidative respiration to denitrification when soil conditions are anoxic (Sanford *et al.*, 2012). Non-denitrifying N₂O reducers are denitrifiers with genes for N₂O reduction only with atypical NosZ gene which encodes for N₂O reductase (N₂OR) and are more diverse and tolerate a wider range of conditions such as anoxic, microaerophilic, oxic, psychrophilic, piezophilic, thermophilic, and halophilic (Sanford *et al.*, 2012). All denitrifiers in natural environments are assumed to be capable of complete denitrification resulting in N₂ as the final product (Firestone and Davidson, 1989). Bacteria and archaea containing the N₂OR also have the potential to convert N₂O to N₂ (Jones *et al.*, 2011).

2.3 Atmospheric N₂O fluxes from boreal soils

Microbial activity and abiotic environmental factors are important prerequisites for the efficiency of the N cycle in most terrestrial ecosystems, especially in N-limited ecosystems. N₂O fluxes from boreal forests are generally far less than those of tropical ecosystems and have perceived capabilities to have a much greater potential in these regions to act as a sink for N₂O (Kroeze *et al.*, 2007). In a study conducted in eastern Finland on afforested organic agricultural soils and organic agricultural soils in active use, the measured N₂O flux values were in the range of 100-2500 μg N₂O-N m⁻² h⁻¹ with intermittent uptakes at sites with high water table level (Maljanen *et al.*, 2012). This is probably due to the fact that the boreal forests are in regions of low temperatures and produce little N₂O with emission rates less than 0.10 Tg N per year (Zhuang *et al.*, 2012). However, in boreal soils from southern Russia and Canada N₂O emissions have been found to be relatively high at rates above 0.20 kg N₂O-N ha⁻¹ yr⁻¹ and this is primarily due to high soil organic matter content and moist climate (Zhuang *et al.*, 2012).

Some studies have also shown N₂O uptake in soils in the boreal region. Net N₂O uptakes of up to $-77\mu\text{g N}_2\text{O-N m}^{-2} \text{ h}^{-1}$ has been measured from drained and abandoned, drained and afforested and active peat extractions in Finland. This uptake has been observed when soil pH and mean soil C:N ratio ranged between 3.9-5.9 and 17.6-24.2 respectively; with higher N₂O uptake in soils with highly saturated water tables (Maljanen *et al.*, 2012). This is because N-limited and high moisture ecosystems such as boreal mires and pristine boreal peatlands support N₂O uptake (Chapuis-Lardy *et al.*, 2007). It is however important to note that there are huge uncertainties involving boreal N₂O fluxes due to high spatial and temporal variability (Ambus *et al.*, 2006; Kellmann and Kavanaugh, 2008).

The highest spatial variability in soil N₂O fluxes is always found in areas known as hot spots (Savage *et al.*, 2014). “Hot spots” are areas that display disproportionately high reaction rates of biogeochemical activity relative to the surrounding area (McClain *et al.*, 2003). Hot spots are often chemically limited because one reactant is unstable in the dominant biogeochemical environment, or because the reaction only proceeds under particular/specific conditions (for example, anoxia) (McClain *et al.*, 2003). Biogeochemical activity can be further categorized as “hot” in the temporal dimension also known as “hot moments” whereby there are periods of intense biogeochemical reactions for a short period of time relative to longer intervening time periods. Hot moments can be delineated at time scales ranging from instants to millennia (McClain *et al.*, 2003). For example, McClain *et al.*, (2003) reported that denitrification hot spots within unsaturated soil profiles become active during hot moments in unsaturated zones where hydrological flow paths are intermittent with strong seasonal variations.

Denitrification hot spots have been identified across spatial scales range from soil profiles to larger basins. In a soil profile of the scale 1-10m, denitrification hot spots have been observed to occur around patches of labile organic matter, for example, plant detritus or manure (Christensen *et al.*, 1990; Petersen *et al.*, 1996); at the anaerobic center of large soil aggregates (Seech and Beauchamp 1988; Højberg *et al.*, 1994); or in earthworm casts (Svensson *et al.*, 1986; Parkin and Berry 1994). These hot spots occur because of the movement of nitrate-rich water into an organic-reducing substrate (Parkin, 1987; Johnston *et al.*, 2001). Consequently denitrification hot spots require oxic-anoxic interfaces meeting a continual water flow, whereby oxic conditions are needed for nitrification to occur producing nitrate and anoxic conditions needed for denitrification and water acting as a transport medium (McClain *et al.*, 2003).

2.4 Factors affecting N₂O sinks

There are several physio-chemical factors affecting the potential for soils to serve as N₂O sinks. These include soil oxygen, soil pH, soil temperature, nutrient availability, soil moisture content and soil type.

2.4.1 Effects of soil oxygen on N₂O fluxes

Soil oxygen is important in controlling N turnover through the processes of nitrification and/or denitrification (Li *et al.*, 2000 and Schurgers *et al.*, 2006). Aerobic soils are generally sources of N₂O but small uptake rates have sometimes been observed in dry soils (Duxbury and Moiser, 1993) and wet grass pastures (Ryden 1981, 1983). In anaerobic soils, a higher possibility for complete denitrification from N₂O to N₂ occurs. Therefore anaerobic soils are generally assumed to be the main potential sinks for N₂O (Erich *et al.*, 1984). Despite this assumption, there has been no large, constant N₂O uptake observed in soils. For example, in flooded rice fields N₂O uptake has been largely dependent on the time of cropping seasons (Minani and Fukushi, 1984 and Parashar *et al.*, 1991). The observed small N₂O uptake in flooded rice fields can be attributed to slow rates of dissolution and transport of atmospheric N₂O in wet soils and hence prevent these sites from being a significant regulator of N₂O emissions (Mosier *et al.*, 1998).

In most surface soils, particularly in fertilized soils containing sufficient organic carbon and nitrate, the presence of oxygen most commonly limits denitrification (Firestone and Davidson, 1989). However, denitrification is extremely variable in space and time making it difficult to accurately ascertain rates of denitrification and N₂O exchange in the field (Lapitan *et al.*, 1999; Hofstra and Bouwman, 2005). N₂O was found to be produced and consumed in both aerobic and anaerobic conditions when oxygen concentrations in soils were primarily affected by soil moisture content (Martikainen *et al.*, 1993; Aerts and Ludwig, 1997). In oxygen-limited conditions, N₂O becomes the sole electron acceptor for denitrifying microbes thus altering N₂O fluxes in soils (Butterbach-Bahl *et al.*, 2013).

2.4.2 Effects of soil pH on N₂O fluxes

Under field conditions, denitrification rates have been found to be lower in acidic soils compared to neutral or slightly alkaline soils (Kroeze *et al.*, 2007). Simek and Cooper (2002) have described the amount of N₂O relative to N₂ formed during denitrification as pH dependent, mostly decreasing when pH increases. Hence, it can be deduced that the highest rates of N₂O reduction occurs in neutral to alkaline soils. This could be explained by N₂O reductase being hindered by a low pH (Richardson *et al.*, 2009).

Moreover, soil pH is a major factor in the process of nitrification either directly or through its effects on soil cation exchange capacity (Robertson, 1989). Optimum pH values for nitrification are noted to range from 6.5 to 8 (Simek and Copper, 2002). When nitrate production increases through nitrification in soils at neutral pH, it could lead to a limited reduction of atmospheric N₂O (Kroeze *et al.*, 2007). According to Maljanen *et al.*, (2012), sites with slightly higher mean soil pH (4.9) produced lower N₂O than soils with slightly lower mean soil pH (4.4)

2.4.3 Effect of soil temperature on N₂O fluxes in natural ecosystems

Soil temperature is a significant factor affecting denitrification which is one of the major processes affecting N₂O fluxes. Typical temperature coefficient (Q₁₀) values for denitrification range from 5 to 16 (Ryden, 1983) and the ratio of N₂O to N₂ produced decreases with increasing temperature (Firestone and Davidson, 1989). Sommerfeld *et al.*, (1993) observed that N₂O emissions were high during the warm growing season in a temperate climate, and this was attributed to high microbial activity. According to Butterbach-Bahl (2013), many microbial processes in the nitrogen cycle are temperature-sensitive. Hence when temperature increases, soil respiration increases leading to decreased soil oxygen concentrations and increased soil anaerobiosis. Therefore, it might be possible that the potential for N₂O uptake through denitrification increases at higher temperatures. However, there have also been some contradictory results showing high N₂O emissions at low soil temperatures during the winter and also during freezing and thawing events (Papen and Butterbach-Bahl, 1999; Maljanen *et al.*, 2007).

2.4.4 Effect of nutrient availability on N₂O fluxes

N-limitation in soils has been shown to cause intermittent N₂O uptake (Rosenkranz *et al.*, 2006; Vanitchung *et al.*, 2011). In an acacia reforestation site with *Acacia mangium* trees, emission of N₂O was observed possibly due to the N-fixation activity of the trees which provided extra nitrogen to the soil (Vanitchung *et al.*, 2011). Higher N₂O emissions have also be observed in nitrogen fertilized cropping systems and also in tree-based systems where litter fall provide additional nutrient availability (Palm *et al.*, 2002).

Atmospheric nitrogen deposition has been found to positively correlate with N₂O emissions (Butterbach-Bahl *et al.*, 1998 and Zechmeister-Boltenstern *et al.*, 2002). Even minor increases in deposition rates over extended durations may lead to changes in the nitrogen cycle of sensitive ecosystems and affects N₂O sink potentials in soils (Bobbink *et al.*, 1998; Bouwman *et al.*, 2002). This is largely because nitrogen enrichment of ecosystems partly

suppresses nitrogen limitation resulting in a reduced N₂O sink strength. According to Kroeze *et al.*, (2007) even when sink activity is seasonal, this reduction of sinks of N₂O may be important considering the extensive areas of nitrogen-affected ecosystems which may in turn lead to increasing atmospheric N₂O concentrations. Therefore, the total availability of nitrogen can also be a major driver of N₂O soil emissions (Butterbach-Bahl *et al.*, 2013).

High nitrate availability has been usually associated with high N₂O emissions (Maljanen *et al.*, 2012). In temperate regions, substrate accumulation in small water films in soils has also been known to result in high N₂O emissions (Papen and Butterbach-Bahl, 1999; Teepe *et al.*, 2000). Organic soils rich in C substrates have been found to be a significant source of N₂O emissions in boreal regions (Alm *et al.*, 2007). Blicher-Mathiesen and Hoffmann (1999) have noted that high nitrate concentrations commonly inhibit N₂O reductase activity which elucidates the strong correlation between nitrate availability and N₂O build-up observed in soils (Skiba *et al.*, 1998). This supports the observation of net uptake of N₂O in grassland and forest soils where nitrate concentrations are typically low (e.g. <1-2 µg NO₃⁻ B/g soil; Butterbach-Bahl *et al.*, 1997). Nitrate concentrations are predicted to be lowest in environments with limited use of N-fertilizers and/or high plant uptake of nitrogen and also where nitrification does not occur (e.g. lack of oxygen and ammonia) (Kroeze *et al.*, 2007). Hence, this results in limited amount of electron acceptors in these environments and N₂ will be predominantly evolved. Moreover, in soils with an absence of oxygen, adequate organic carbon is present to support N₂O reduction (International Plant Nutrition Institute, 2016).

2.4.5 Effects of soil moisture content

One critical factor that affects boreal N₂O fluxes is soil moisture content. This can be assessed differently e.g. as precipitation (Werner *et al.*, 2007), water-filled pore space (WFPS) (Davidson *et al.*, 2000) or water table level (WTL) (Maljanen *et al.*, 2012). At above 60% WFPS denitrification will occur because there is no absolute anaerobic situation and N₂O can be produced as a by-product but when WFPS percentages are higher, the anaerobic situation is more pronounced and the production of N₂O will decrease (Davidson *et al.*, 2000).

Studies in boreal ecosystems showed that raising the water level close to the soil surface is likely to reduce N₂O emissions with N₂O emissions possibly mitigated when WTL is lower than 70cm (Maljanen *et al.*, 2012). Thus, it can be deduced that soil moisture content is closely coupled with soil microbial activity which consequently affects nitrification and denitrification determining N₂O fluxes. High soil moisture content results in anoxic conditions which amplify N₂O production through denitrification (Vanitchung *et al.*, 2011).

Soil moisture content also exerts considerable influence on activities of soil microbes, delivery of electron donors (NH_4^+ , DOC) and electron acceptors (O_2 , NO_3^-) and in the diffusion of N trace gases from soils (Firestone and Davidson, 1989). It also regulates the availability of oxygen to soil microbes; and high water level in the soil enables high microbial N turnover rates ensuring that there are substrates available for soil microbes (Goldberg *et al.*, 2010; Butterbach-Bahl *et al.*, 2013).

In addition, WFPS might be linked with nitrate concentrations in affecting N_2O emissions. Limited nitrate conditions and higher WFPS stimulate denitrification enabling denitrifiers to utilize N_2O as an electron acceptor and inhibiting nitrification (Vanitchung *et al.*, 2011). Moreover, increased N substrates and easily degradable C availability have also been found to increase microbial N_2O emissions (Papen and Butterbach-Bahl, 1999).

2.4.6 Effect of soil type on N_2O fluxes

2.4.6.1 Soil Carbon-to-Nitrogen ratio (C:N ratio)

Soil carbon-to-nitrogen ratio (C:N ratio) determines the decomposability of soil organic matter which in turn has a critical impact on soil nitrogen availability. Relationships between soil C:N might play a key role in nitrous oxide emissions. Soil residues with lower C:N ratios have been observed to have a higher decomposition rate hence providing more dissolved organic carbon (DOC) and therefore increasing N_2O emissions (Huang *et al.*, 2004). Soil C:N ratio affects the mineralization of plant residues and consequently N_2O emissions (Aulakh *et al.*, 1991 and Németh *et al.*, 1996). Organic amendments to a well-aerated soil which decrease C:N ratios have been shown to increase N_2O emissions (Bremmer and Blackmer, 1981). Substrates with C:N ratios <20 have a higher decomposition rate resulting in ammonium release via mineralization increasing N_2O emissions. Substrates with ratios of 25-75 also decompose quickly but N mineralization is inhibited by increased microbial immobilization and protein complexation by polyphenols when the cells lyse. In a modelling study in Scandinavia and the Baltic States, soils with elevated C stocks had considerably higher N_2O emissions than ambient soils; with values of $>0.75 \text{ kg N ha}^{-1}$ being calculated (Kesik *et al.*, 2005). High N_2O emissions were also predicted via modelling for soils with high amounts of organic carbon content in the forest floor in Southwest Finland and in the Northern parts of Sweden (1.0 to $1.8 \text{ kg N ha}^{-1} \text{ yr}^{-1}$) (Kesik *et al.*, 2005).

2.4.6.2 N₂O emissions from Organic vs mineral soil types

N₂O emissions from mineral soils in boreal forests are in the range of 0.1 and 0.3 kg N ha⁻¹ yr⁻¹ (Brumme *et al.*, 2005), while forest soils, rich in organic matter, emitted N₂O in the range of 1.0 to 10.0 kg N ha⁻¹ yr⁻¹ (Maljanen *et al.*, 2001, 2003; von Arnold *et al.*, 2005). N₂O emissions from peat soils (organic soil) that have been used for agriculture prior to forestation emitted the most N₂O (Maljanen *et al.*, 2003). Therefore, it can be concluded that large N₂O emissions occur as a result of drainage and cultivation of organic soils due to enhanced mineralization of old, N-rich organic matter (Guthrie and Duxbury, 1978; Martikainen *et al.*, 1996; Velthof *et al.*, 1996). Nutrient poor organic forests were noted to emit negligible amounts of N₂O (Regina *et al.*, 1996). However, drained organic soils with no fertilizer additions were noted to show much higher emissions of N₂O of up to 100 kg N- N₂O ha⁻¹ yr⁻¹ than mineral soils (Regina *et al.*, 1996).

2.4.6.3 Natural vs altered ecosystems

Human activities like agriculture affect the nitrogen cycle by increasing N₂O emissions. The production of synthetic nitrogen fertilizers and cultivation of N₂-fixing plants play a key role in steadily increasing nitrogen into the biosphere thus altering the nitrogen cycle (Vitousek *et al.*, 1997). Nitrogen fertilizers that contain ammonium and nitrate have been known to elevate the emissions of N₂O immediately after addition (Eichner, 1990, Chang *et al.*, 1998). Drainage along with other agricultural practices such as ploughing, liming and fertilization contribute to higher pH values in soil and stimulate decomposition of N-rich organic matter (Maljanen *et al.*, 2012) and nitrogen mineralization (Freeman *et al.*, 1996).

It is known that emissions of N₂O from pristine peatlands are negligible but in drained peatlands there is an increase in N mineralization leading to greater emissions of N₂O due to nitrification and denitrification (Regina *et al.*, 2004). In Nordic countries, agricultural activities conducted in organic soils resulted in N₂O emissions on average four times higher than those from mineral soils, indicating that N₂O derived from soil organic carbon decomposition dominates overall fluxes (Mu *et al.*, 2014). Moreover, in organic soils the variability of soil C/N ratio may be one of the dominant factors determining N₂O emissions in organic soils (Mu *et al.*, 2014).

Several studies have noted that clayey and highly fertile soils result in higher N₂O emission (Matson and Vitousek, 1987; Verchot *et al.*, 1999). Forest soils are more inclined to provide acidic conditions which may select microorganisms to different processes according to their tolerance of pH ranges (Muñoz *et al.*, 2010). Acidic soil conditions of coniferous forests in

Western Europe contain heterotrophic bacteria and fungi promoting nitrification with the bacteria *Arthrobacter sp.* seemingly most highly adapted to initiate heterotrophic nitrification (Brierley and Wood, 2001).

In the forests of South-Central Chile, dominated by volcanic soils the N-cycle is extremely efficient resulting in low productions of N gases to the atmosphere due mainly to the physicochemical characteristics of these soils (Chorover, 2002; Godoy *et al.*, 2003). In pristine soils, N conservation can be attributed to consumption by microorganisms and vascular plants by net primary production (Muñoz *et al.*, 2010). There is a scarce production of nitrate in these soils resulting in nitrification requiring an extra-consumption of energy at the ecosystem level (Huygens *et al.*, 2008). Hence, this results in an increased amounts of N immobilized by microbial action and adsorption of inorganic forms of N onto clay colloid surfaces (Bengtsson and Bergwall, 2000; Huygens *et al.*, 2008). It has also been noted that land use change from native forest to forest plantations and grassland remarkably increased N mineralization and nitrification in soils of New Zealand (Parfitt *et al.*, 2003).

2.4.7 Hole in the pipe model

Firestone and Davidson (1989) have proposed a model known as “hole-in-the-pipe” (HIP) which explains how microbiological and ecological factors affected soil emission of NO and N₂O fluxes. Several studies have indicated that nitrogen fertilization encouraged production of one or both gases (Williams *et al.*, 1992). On the contrary, in unfertilized soils net nitrogen mineralization and net nitrification have been found to positively correlate with N₂O emissions (Robertson and Tiedje, 1984, Matson and Vitousek, 1987). In the HIP model, the total production of NO and N₂O is assumed to be directly linked to the availability of nitrogen in the soil. The HIP model also attempts to link soil water content to the ratios of N₂O:NO emissions as a function of soil water content (Davidson *et al.*, 2000).

The HIP model is expressed by leaky pipes representing two major processes nitrification and denitrification. The rate of flow of nitrogen through these pipes is comparable to the rates of nitrification and denitrification and shows nitrogen cycling through the ecosystem (Davidson *et al.*, 2000). “Holes” in the pipe represent trace gases of NO and N₂O leaking out of the pipe and the sizes of these holes correspond primarily to soil water content (Davidson *et al.*, 2000).

3. Materials and Methods

3.1 Study site

The first study site was located in eastern Finland in Heinävesi (62°26'N, 28° 38'E) which is an upland forest of Myrtillus type (MT) (Hotanen *et al.*, 2008). The main tree species located at the site are spruce (*Picea abies*), pine (*Pinus sylvestris*) and birch (*Betula pendula* and *Betula pubescens*) with a stand age of 80 years and the dominant soil type in this area can be characterized by a humus layer and mineral soil.

The second study site was located in eastern Finland in Salmisuo, Ilomantsi (62°47'N, 30°56'E) in a low-sedge *Sphagnum papillosum* pine fen. The Salmisuo peatland complex is an eccentric bog with some minerotrophic strips. Plant species indicate average water tables and nutritional statuses of the mire types and microsites. Flarks are the wettest microsites, with an average water table depth from 0 to –5 cm while lawns have an average water level depth 5–20 cm below the peat surface. Hummocks rise above their surroundings and have an average water table over 20 cm below the peat surface. The vegetation in Salmisuo enabled all three distinctive forms - hummocks, lawns and flarks, to be recognized in the low-sedge *S. papillosum* pine fen.

The proportion of different microsites were determined by evaluating their coverage in 20 vegetation squares (1×1 m, along a transect with 5-m intervals). The most typical hummock species was *Sphagnum fuscum* (Schimp.) Klinggr., with *S. angustifolium* (Russow) C. Jens. and *Eriophorum vaginatum* L. also found in the lower parts of the hummocks. The lawn moss layer was dominated by *S. angustifolium* and *S. balticum* (Russow) C. Jens., accompanied by some *S. magellanicum* Brid. and *S. papillosum* Lindb. The field layer in the lawns consisted mainly of *E. vaginatum* with *Andromeda polifolia* L., *Vaccinium oxycoccos* L. and *Carex pauciflora* Light. The major mosses in the flarks were *S. majus* (Russow) C. Jens., *S. balticum* and *S. angustifolium*, and the only vascular plants on flarks were *Scheuchzeria palustris* L. and *E. vaginatum*. Between the mineral soil and the low-sedge *S. papillosum* pine fen there was a narrow tall-sedge fen lag consisting of a lawn vegetation in which the major moss species was *S. angustifolium* and the field layer was dominated by *C. rostrata* Stokes. In this study, upper peat layers of 0-10cm and lower peat layers of 10-20cm were sampled.

Organic and mineral soil at the Heinävesi site (Fig. 2) and profiles of 0-10cm depth and 10-20cm depth at the Salmisuo site were collected on the 2nd of June 2014 and the 8th of July

2014 respectively. Soils were collected and sealed in ziplock bags and refrigerated in cold room of temperatures up to 4°C until physical and chemical analysis of soils were conducted. Soils were then homogenized manually by picking out roots and debris. Soil pH was measured from soil-water suspension (1:2-3 v/v) by using a pH meter (WTW pH-Electrode SenTix® 81, Germany) while electrical conductivity was measured using an EC meter (WTW TetraCon® 325, Germany). Soil organic matter was also analyzed for the respective soil layers utilizing the weight loss on ignition method (Reeuwijk, 2002). Samples were first oven-dried at 105°C in a muffle furnace and thereafter the samples were burnt at 450°C (Reeuwijk, 2002). The weight loss that occurred at this temperature was then correlated to the soil moisture content.



Figure 2: Organic and mineral layers of forest soil at Heinävesi site

3.2 Experimental set-up

3.2.1 Effect of oxygen on N_2O , NH_4^+ and NO_3^- concentrations

The experiment was carried out in flask-microcosms (Figure 3) in oxic and anoxic conditions. Sacrificial sampling was carried out during the experiment. To ensure that there was no N_2O consuming activity already present, pre-incubation was done for 5 days whereby bottles were degassed and flushed with 100% helium. This degassing and flushing was done twice to ensure that there was complete removal of accumulated N_2O during the 5 days. After the pre-incubation period, N_2O consumption was activated in the respective soil layers (mineral/organic; upper 0-10cm/lower 10-20cm) of the Heinävesi and Salmisuo soils with

addition of 15-N N₂O (98% atm, 1500 ppb) to the headspace of the bottles. The respective soil layers of each soil site were then subjected to oxic and anoxic conditions. Oxic conditions were achieved by adding 21% volume of O₂ to the headspace of the bottles while anoxic conditions did not have any additions of O₂. Sampling of nitrous oxide gas, nitrate and ammonium was then done at various time points for each site (0, 3, 12, 48, 72, 168 hours for Heinävesi and 0, 3, 24, 48, 72, 168 hours for Salmisuo). There were five replicates for each treatment (anoxic/oxic conditions) and incubation experiments were conducted at room temperatures of 22°C.



Figure 3: Flask-microcosm experiment (Peat samples)

N₂O concentrations were then determined by Gas Chromatography analysis (Hewlett Packard 5890 Series II, U.S.A). Ammonium-N (NH₄⁺-N) and nitrate-N (NO₃⁻-N) were extracted from integrated soil samples with 0.5M K₂SO₄ (soil:K₂SO₄ 1:2 v/v, 175 rev min⁻¹, 1 hour) at various time points during the incubation period (168h). The extracts were filtered (Blauband 589³ BlueRibbon filter paper) overnight and stored at -20°C until analyzed for NH₄⁺ and NO₃⁻. The concentration of NH₄⁺ and NO₃⁻ in the extracts were measured by UV-Visible spectrophotometry (Philips PU 87501 UV/VIS) using microtiter plate format and following the protocol of Fawcett and Scott (1960) for NH₄⁺ (630 nm) and Griess method for NO₃⁻ in aqueous solution (544 nm) (Miranda *et al.*,2001).

3.2.2 Effect of electron donors on mineral forest soil and lower 10-20cm peat soil on N₂O concentrations in oxic conditions

We also wanted to understand the effects of electron donors on the N₂O fluxes in mineral layer of Heinävesi and 10-20cm layer of Salmisuo sites. Five electron donors (succinate, butyrate, formate, propionate, lactate) were added to soils (Heinävesi: mineral; Salmisuo: 10-20cm depth). There were 3 replicates of each treatment and 3 controls for each site. Soils were added to bottles and pre-incubated in anoxic conditions for five days at 14°C and degassed and flushed with 100% Helium. N₂O consumption was activated by adding 15-N N₂O (98% atm, 1500 ppb) to the headspace of the bottles. Oxic conditions were then ensured in the bottles by adding 21% volume of O₂ to the headspace of bottles. This was then followed by the addition of 1mM of each electron donor to the respective bottles. Sampling of nitrous oxide was done at various time points (0, 3, 5, 24, 48, 72, and 168 hours) for both soils.

3.3 Statistical Analysis

Statistical analysis was performed with the IBM SPSS Statistics 21. The Shapiro-Wilk test of normality of variables was first carried out on all data to check for normality of data. As the data did not conform to normality, non-parametric tests (Kruskal-Wallis and Mann-Whitney U) were used to compare the data sets between sites over time.

4. Results

4.1 Soil properties of the study sites

Heinävesi forest soils have a considerably lower NH_4^+ content than Salmisuo peatland soils with the lowest NH_4^+ content observed in the mineral layer of Heinävesi soils and the highest NH_4^+ content in the 10-20cm layer of the Salmisuo soil (Table 2). The same trend was observed for the NO_3^- concentrations, with the Heinävesi soils having a notably lower NO_3^- content than the Salmisuo soils. The lowest NO_3^- content was observed for the mineral layer of the Heinävesi soil but the highest NO_3^- content was observed in the upper (0-10cm) layer of the Salmisuo soil. For the Salmisuo soil the upper layer (0-10cm) had nearly twice as much NO_3^- than the lower layer (10-20 cm) (Table 2).

Table 2: Background Soil properties of the study sites

| Site | Soil Type | $\mu\text{g N-NH}_4/\text{g}_{\text{dw}}$ | $\mu\text{g N-NO}_3/\text{g}_{\text{dw}}$ | pH | Temperature | EC ($\mu\text{ S/Cm}$) | Soil Moisture (%) |
|-----------|-----------|---|---|------|-------------|--------------------------|-------------------|
| Heinävesi | Mineral | 2.78 ± 0.90 | 0.023 ± 0.003 | 4.89 | 21.1 | 15 | 8.31 |
| Heinävesi | Organic | 5.30 ± 0.57 | 0.033 ± 0.008 | 4.68 | 21.1 | 14 | 31.71 |
| Salmisuo | 0-10 | 26.78 ± 0.26 | 0.87 ± 0.53 | 4.25 | 22.3 | 20 | 88.59 |
| Salmisuo | 10-20 | 33.62 ± 1.61 | 0.46 ± 0.13 | 4.76 | 22.3 | 18 | 90.76 |

Both soils had very similar pH range and were largely acidic. The upper layer (0-10cm) of the Salmisuo soil was the most acidic, followed by the organic layer of the Heinävesi soil, the lower layer of Salmisuo soil (10-20cm) and the mineral layer of Heinävesi respectively (Table 2). In general, the Salmisuo peat soil had higher soil moisture content than the Heinävesi upland forest soils. Both peat layers (Salmisuo soil) have similar soil moisture contents but the organic upland forest soil (Heinävesi soil) had higher soil moisture content than the mineral layer (Table 2).

4.2 Effects of Anoxic conditions on N₂O fluxes in forest soils (Heinävesi)

When forest soils were exposed to anoxic conditions, N₂O concentrations decreased in both mineral and organic soil layers (Figure 4a). A pronounced decrease in N₂O concentrations in both soil layers was observed between 3 and 48 hours of incubation with the organic soil showing greater mean net consumption of N₂O (decreased in concentration) than the mineral soil throughout the experiment (Figure 4a, Table 3). The effect of anoxia on this decrease in N₂O was statistically significant during this time frame (Table 4). Anoxic conditions also significantly resulted in the near total consumption of the N₂O in the organic layer by the end of the experiment (Figure 4a). The forest mineral layer however, was only able to reduce half of the N₂O concentrations by the end of the experiment compared to initial N₂O fluxes (Figure 4a).

In addition, a statistically significant increase in ammonium concentrations in the organic layer of the forest soil was measured throughout the experiment under anoxic conditions (Figure 4b, Table 5). For the mineral layer, the effect of anoxia was not significant since the concentrations of ammonium remained constant and negligible throughout the experiment (Figure 4b, Table 5). On the other hand, nitrate concentrations in both soil layers (organic and mineral) showed a slightly decreasing trend throughout the experiment (Figure 4c), but it was not statistically significant (Table 6).

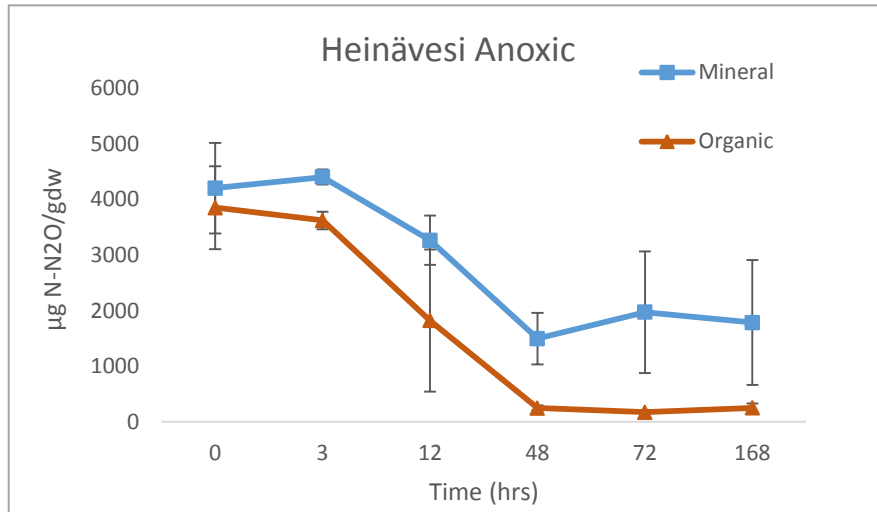


Figure 4a: Effect of anoxic conditions on N₂O fluxes in Heinävesi soils

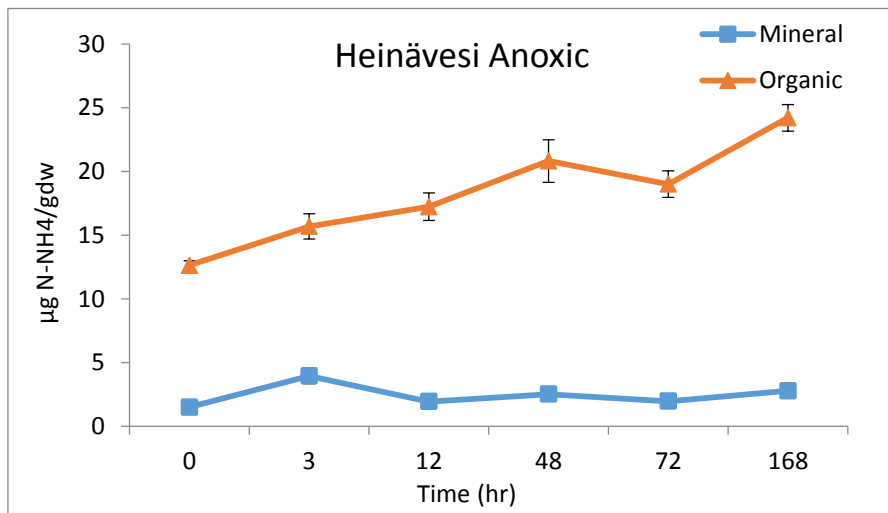


Figure 4b: Effect of anoxic conditions on NH₄⁺ concentrations in Heinävesi soils

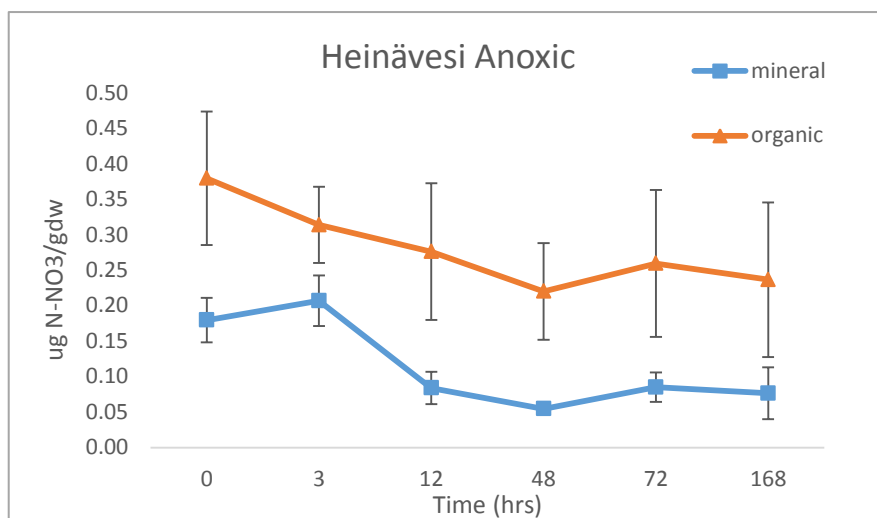


Figure 4c: Effect of anoxic conditions on NO₃⁻ concentrations in Heinävesi soils

Table 3: Net N₂O consumption/production potential in Heinävesi and Salmisuo soils in anoxic/oxic treatment

| Site | Soil Type | Anoxic Treatment | Oxic Treatment |
|-----------|-----------|------------------|----------------|
| Heinävesi | Mineral | -91.28 | 1.10 |
| Heinävesi | Organic | -322.14 | 493.53 |
| Salmisuo | 0-10cm | -187.38 | -40.61 |
| Salmisuo | 10-20cm | -319.06 | -63.15 |

Table 4: Effect of oxygen on nitrous oxide (N₂O-N) concentrations (* indicating p<0.05)

| Site | Soil | Time | Significance |
|-----------|---------|------|--------------|
| Heinavesi | Mineral | 0 | 0.221 |
| | | 3 | 0.016 * |
| | | 12 | 0.009 * |
| | | 48 | 0.009 * |
| | | 72 | 0.047 * |
| | | 168 | 0.117 |
| | Organic | 0 | 0.329 |
| | | 3 | 0.009 * |
| | | 12 | 0.016 * |
| | | 48 | 0.009 * |
| | | 72 | 0.009 * |
| | | 168 | 0.009 * |
| Salmisuo | 0-10cm | 0 | 0.117 |
| | | 3 | 0.016 * |
| | | 24 | 0.009 * |
| | | 48 | 0.009 * |
| | | 72 | 0.009 * |
| | | 168 | 0.009 * |
| | 10-20cm | 0 | 0.047 * |
| | | 3 | 0.009 * |
| | | 24 | 0.009 * |
| | | 48 | 0.009 * |
| | | 72 | 0.009 * |
| | | 168 | 0.009 * |

Table 5: Effect of oxygen on ammonium (NH₄⁺-N) concentrations (* indicating p<0.05)

| Site | Soil | Time | Significance |
|-----------|---------|------|--------------|
| Heinavesi | Mineral | 0 | 0.94 |
| | | 3 | 0.465 |
| | | 12 | 0.068 |
| | | 48 | 0.602 |
| | | 72 | 0.475 |
| | | 168 | 0.076 |
| | Organic | 0 | 0.034 * |
| | | 3 | 0.028 * |
| | | 12 | 0.009 * |
| | | 48 | 0.009 * |
| | | 72 | 0.009 * |
| | | 168 | 0.009 * |
| Salmisuo | 0-10cm | 0 | 0.465 |
| | | 3 | 0.602 |
| | | 24 | 0.221 |
| | | 48 | 0.602 |
| | | 72 | 0.028 |
| | | 168 | 0.034 |
| | 10-20cm | 0 | 0.117 |
| | | 3 | 0.699 |
| | | 24 | 0.602 |
| | | 48 | 0.624 |
| | | 72 | 0.289 |
| | | 168 | 0.053 |

Table 6: Effect of oxygen on nitrate (NO₃⁻-N) concentration (* indicating p<0.05)

| Site | Soil | Time | Significance |
|-----------|---------|------|--------------|
| Heinavesi | Mineral | 0 | 0.086 |
| | | 3 | 0.465 |
| | | 12 | 0.142 |
| | | 48 | 0.064 |
| | | 72 | 0.297 |
| | | 168 | 0.456 |
| | Organic | 0 | 0.329 |
| | | 3 | 0.564 |
| | | 12 | 0.076 |
| | | 48 | 0.717 |
| | | 72 | 0.784 |
| | | 168 | 0.175 |
| Salmisuo | 0-10cm | 0 | 0.251 |
| | | 3 | 0.465 |
| | | 24 | 0.245 |
| | | 48 | 0.472 |
| | | 72 | 0.327 |
| | | 168 | 0.251 |
| | 10-20cm | 0 | 0.754 |
| | | 3 | 0.347 |
| | | 24 | 0.602 |
| | | 48 | 0.251 |
| | | 72 | 0.347 |
| | | 168 | 0.754 |

4.3 Effects of Oxic conditions on N₂O fluxes in forest soils (Heinävesi)

Under oxic conditions, the organic layer showed a gradual and steady increase in N₂O fluxes throughout the experiment with the presence of oxygen significantly increasing N₂O fluxes between the 3rd hour till the end of the experiment (Figure 5a, Table 4). In the mineral layer, N₂O concentrations oscillated around the average value of (5937.5 ± 677.0 μg N-N₂O/gdw) throughout the experiment with a statistically significant increase between the 3rd hour and 12th hour of incubation (Figure 5a, Table, 4). Organic soil showed a mean net production of N₂O while mineral soil had a negligible mean net production of N₂O (Table 3).

Ammonium concentrations in organic layer showed a generally declining trend. There were slight increases in ammonium concentrations in the organic layer between the 12th and 48th hour and also towards the end of the experiment (72 to 168 hours) (Figure 5b). Nitrate concentrations increased steadily throughout the experiment in organic layer but the effect of oxygen was not statistically significant (Figure 5b, Table 6). In mineral layer, there were no distinguishable changes in ammonium concentrations but slightly declining nitrate concentrations throughout the experiment (Figure 5b, 5c). There was also no statistically significant effect of oxygen on ammonium and nitrate concentrations in mineral layer during the experiment (Table 5, 6).

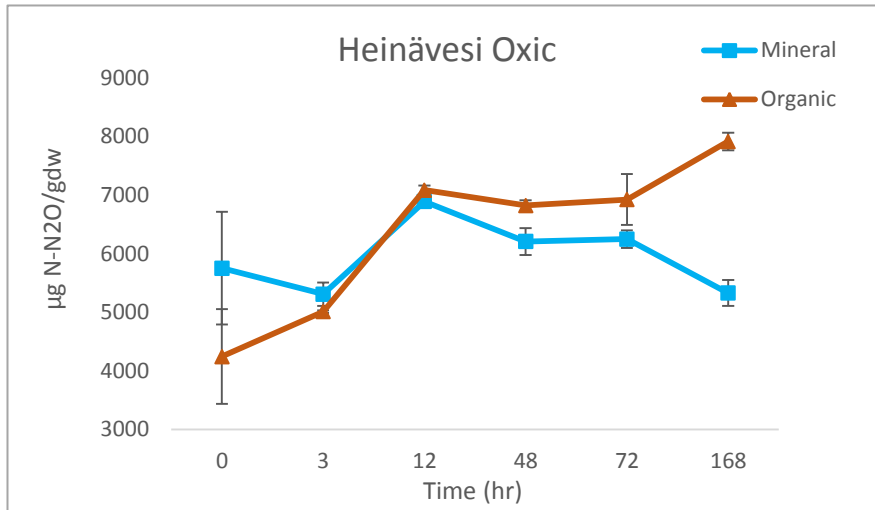


Figure 5a: Effect of oxic conditions on N_2O fluxes in Heinävesi soils

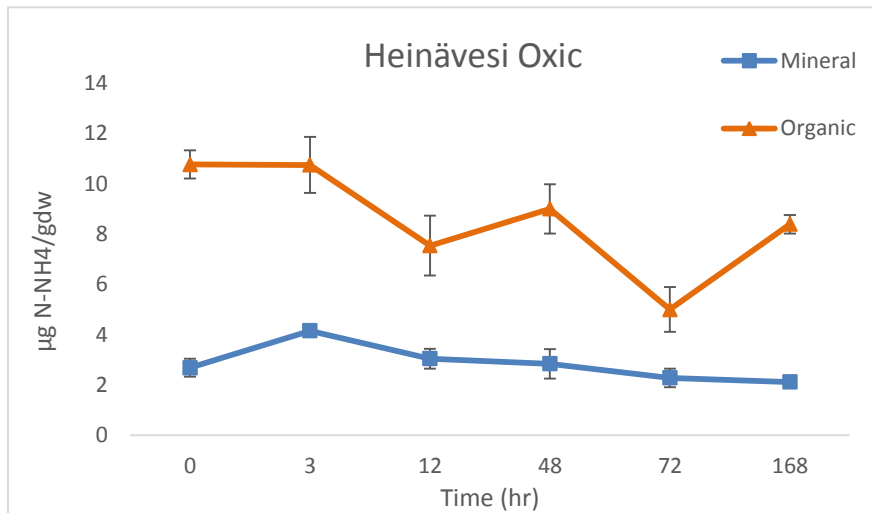


Figure 5b: Effect of oxic conditions on NH_4^+ concentrations in Heinävesi soils

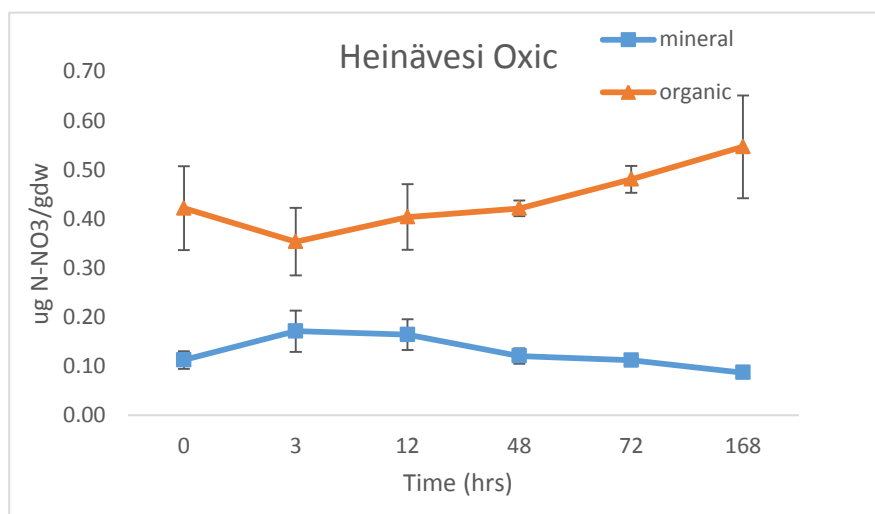


Figure 5c: Effect of oxic conditions on NO_3^- concentrations in Heinävesi soils

4.4 Effects of anoxic conditions on N₂O fluxes in peat (Salmisuo)

Peat soils exposed to anoxic conditions had declining N₂O concentrations throughout the experiment in both layers (Figure 6a). Similar to the forest soils in anoxic conditions (Figure 4a), there was a statistically significant decrease in N₂O fluxes in both peat layers between 3 and 72 hour of incubation (Table 4). The lower peat layer (10-20cm) exhibited greater net consumption of N₂O than the upper peat layer (0-10cm) (Figure 6a, Table 3). Additionally, anoxic conditions significantly resulted in the near total consumption of N₂O in the lower peat layer (10-20cm) by the end of the experiment (Figure 6a). In the upper peat layer (0-10cm) N₂O was not completely consumed but its concentration was roughly 7 times lower compared to the initial N₂O concentration (Figure 6a).

At anoxic conditions, ammonium concentrations in the upper peat layer (0-10cm) remained fairly steady but peaked at the 48th time point of incubation and then subsequently declined till the 72nd hour (Figure 6b). Nitrate concentrations in the upper peat layer (0-10cm) remained consistently low throughout the experiment (Figure 6c). In the lower peat layer (10-20cm), ammonium concentrations fluctuated between a minimum of 9.86µg N-NH₄/gdw and a maximum of 17.22µg N-NH₄/gdw throughout the experiment (Figure 6b). Nitrate concentrations in the bottom peat layer (10-20cm) also showed a rapid decline in the first 3 hours and then declined steadily to the end of the experiment (Figure 6c). The effect of anoxia on the concentrations of ammonium and nitrate were not statistically significant in either soil layer (Table 5, 6).

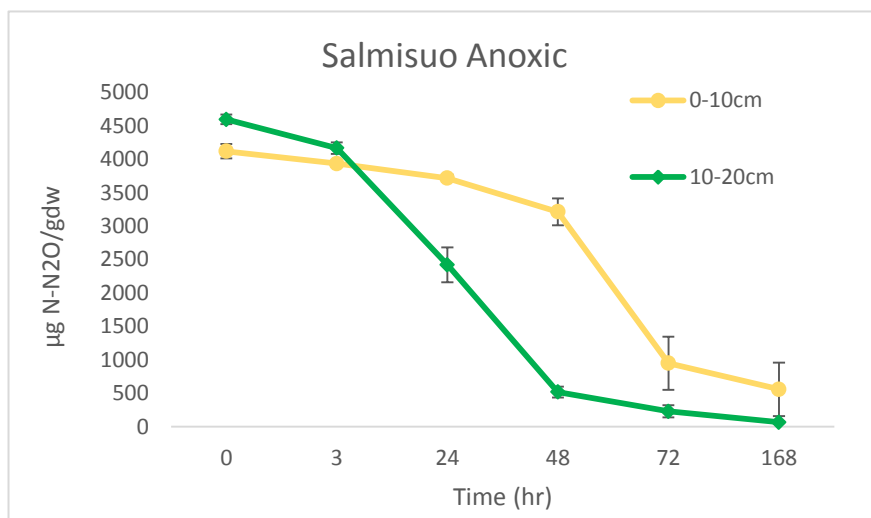


Figure 6a: Effect of anoxic conditions on N₂O fluxes in Salmisuo peat

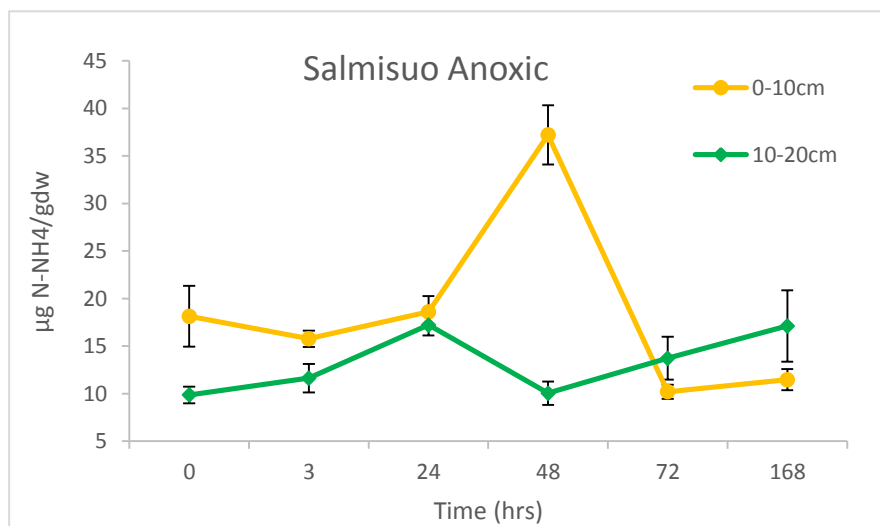


Figure 6b: Effect of anoxic conditions on NH₄⁺ concentrations in Salmisuo peat

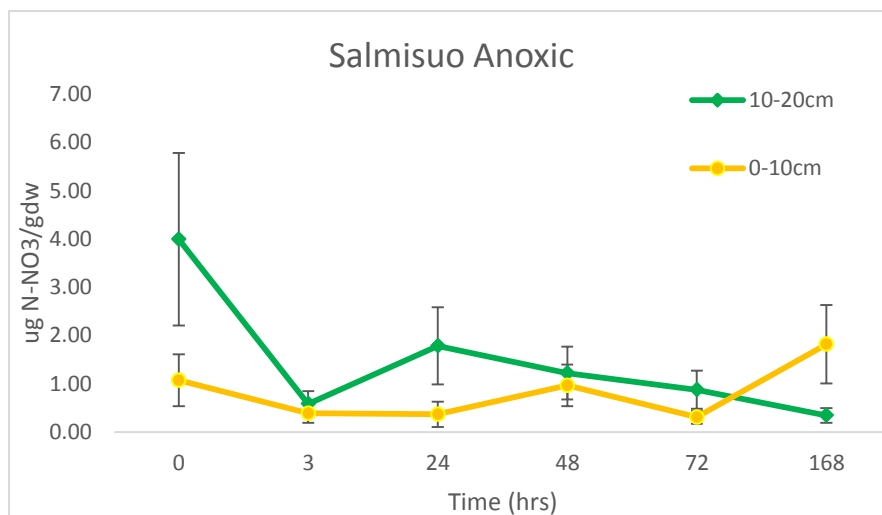


Figure 6c: Effect of anoxic conditions on NO₃⁻ concentrations in Salmisuo peat

4.5 Effects of Oxic conditions on N₂O concentrations in peat (Salmisuo)

In the upper peat layer (0-10cm) N₂O concentrations remained fairly steady in the first 72 hours of incubation while in the lower peat layer, (10-20cm) N₂O concentrations declined slowly between 0 and 72 hours. After the 72nd hour time point, N₂O concentrations in both peat layers declined abruptly till the end of the experiment (Figure 7a). The lower peat layer (10-20cm) averaged a higher consumption potential than the upper peat layer (0-10cm) (Table 3). The effect of oxic treatment on N₂O concentrations of both peat layers was statistically significant throughout the entire experiment with exception for the upper peat layer (0-10cm) at the beginning of the experiment (Table 4).

Ammonium concentrations in the lower peat layer (10-20cm) fluctuated between a minimum of 5.03µg N-NH₄/gdw and a maximum of 46.09µg N-NH₄/gdw throughout the experiment (Figure 7b) while nitrate concentrations increased for the first 24 hours and then decreased steadily to end of the incubation period (Figure 7c). In the upper peat layer (0-10cm), ammonium concentrations increased between 0 and 48 hours and then steadily decreased until the 72nd hour before remaining constant to the end of the experiment (Figure 7b). Nitrate concentrations in upper peat layer (0-10cm) remained fairly constant throughout the experiment (Figure 7c). The effect of oxygen on ammonium and nitrate concentrations was not statistically significant throughout the experiment for either peat layer (Table 5, 6).

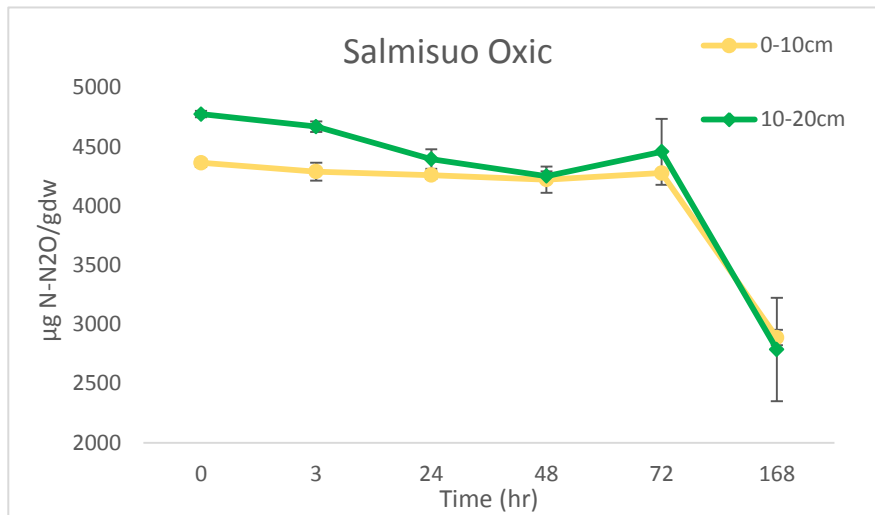


Figure 7a: Effect of oxic conditions on N₂O fluxes in Salmisuo peat

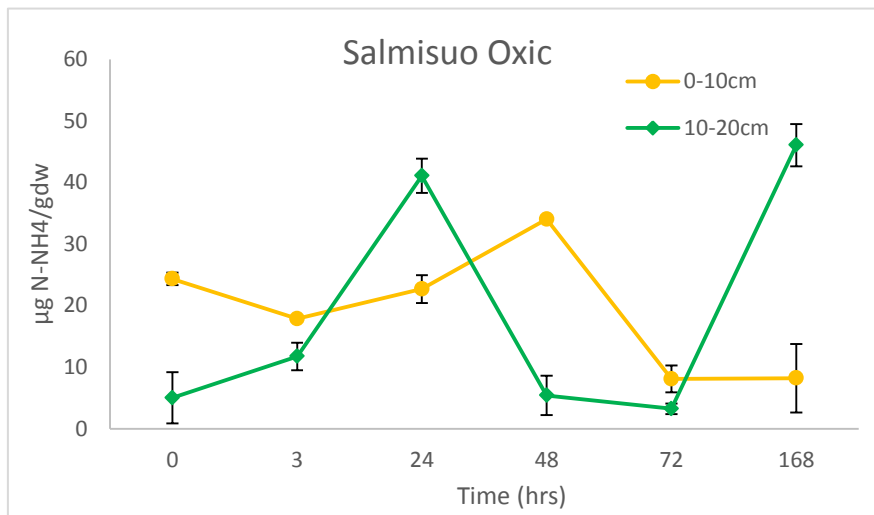


Figure 7b: Effect of oxic conditions on NH₄⁺ concentrations in Salmisuo peat

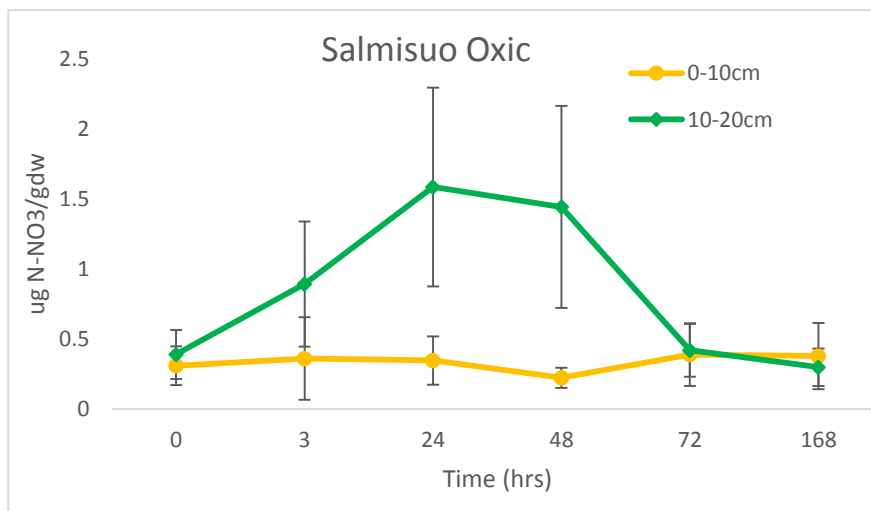


Figure 7c: Effect of oxic conditions on NO₃⁺ concentrations in Salmisuo peat

4.6 Effects of different electron donors on N₂O fluxes from the mineral layer of Heinävesi forest and the lower peat layer (10-20cm) of Salmisuo

Different carbon sources resulted in different net N₂O flux rates in mineral (Heinävesi forest) and peat soils (Salmisuo peat soil) (Figures 8a and 8b). Both soils showed a decline in net N₂O flux rates in the incubations with propionate, indicating the preference for this carbon source during N₂O consumption processes. However, the peat soil (10-20cm layer) from Salmisuo soils exhibited a comparatively larger net N₂O consumption potential with propionate than the mineral forest soil from Heinävesi (see scale in the figures 8a and 8b). The incubations with butyrate and lactate exhibited net N₂O production potential in both soils but the peat soil (10-20cm layer) showed a larger net N₂O production potential. The addition of succinate resulted in a near negligible consumption of N₂O in mineral soils but a considerably greater net N₂O consumption potential in the peat soil (10-20cm layer). In the incubations with formate contrasting effects on net N₂O fluxes were observed with net N₂O production in mineral soil and net N₂O consumption in peat (10-20cm layer).

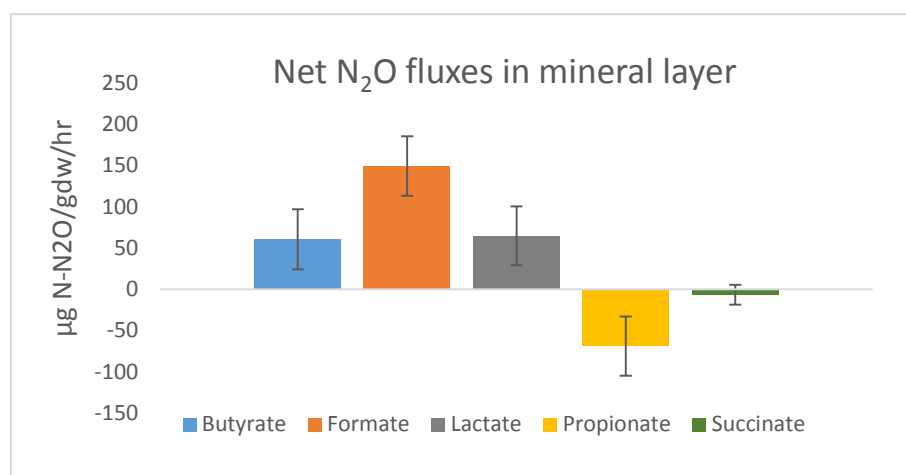


Figure 8a: Effect of different electron donors on net N₂O fluxes in mineral soil

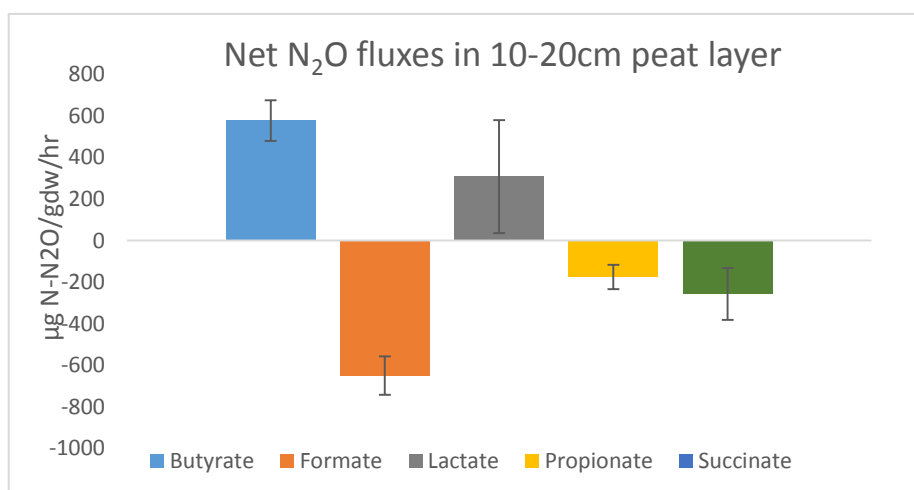


Figure 8b: Effect of different electron donors on net N₂O fluxes in 10-20cm peat

5. Discussion

5.1 Effects of anoxic conditions on N₂O fluxes in forest soils (Heinävesi)

Denitrification is a significant process regulating the nitrogen cycle. In prevailing anoxic conditions, like the conditions in our experiment, the dominant process noted to be able to consume N₂O is denitrification (Conrad, 2002). The results of the present study support these observations because there was a pronounced decline of N₂O concentrations within the first 48 hours of the soil incubation under anoxic conditions for both soil layers. Denitrification also produces N₂O as an intermediate (Butterbach-Bahl *et al.*, 2003; Voss *et al.*, 2013; Ward, 2013) and this probably explains the initial concentrations of N₂O in both soil layers. The differences observed in N₂O consumption rates between the soil layers could be attributed to the differences in physico-chemical properties of each soil layer.

In organic soils, anoxic conditions could have been intensified by the increasing ammonium concentrations during anaerobic decomposition of organic matter (Liikanen, 2003; Ward, 2013). The conditions for complete denitrification could then be built up in the organic soil, resulting in lower N₂O fluxes over time and led to nearly all the N₂O fluxes in the organic layer to be consumed by the end of the experiment. The decreasing nitrate concentrations over time observed in organic soils implies a lack of electron acceptors and supports the notion that N₂ was mainly produced instead of N₂O in these soils. Overall, the results suggested that the organic layer from the forest soil (Heinävesi soils) has a high potential as a sink for N₂O (Erich *et al.*, 1984).

In the mineral soils, there was also a net decrease in N₂O fluxes over time but the process of denitrification did not seem as acute as in the organic layer. This could be due to the soil properties of the mineral layer which contain relatively low ammonium and nitrate concentrations. Several studies have shown that nitrogen limited conditions support N₂O consumption (Flechard *et al.*, 2005; Rosenkranz *et al.*, 2006; Vanitchung *et al.*, 2011). Moreover relatively low nitrate concentrations, like the concentrations measured in the mineral layer from Heinävesi soils (<1µg/gdw soil) throughout the experiment, have also been noted to support N₂O uptake because nitrate is a limiting factor affecting denitrification (Seitzinger, 1988, Butterbach-Bahl, 1997).

The net greater N₂O consumption potential observed in the organic soil layer compared to the mineral soil layer of Heinävesi forest soil has also been described previously for an N-limited Mediterranean pine forest study (Rosenkranz *et al.*, 2006). In that study, the net greater N₂O

consumption potential observed in the organic soil has been attributed to the higher soil moisture content of the organic compared to the mineral soil which intensifies the process of anaerobiosis (Kent *et al.*, 2002). This consequently increases the activity of complete denitrifiers containing NosZ gene resulting in a comparatively higher rate of N₂O consumption in these soils (Kang *et al.*, 2007; Wolf *et al.*, 2010).

5.2 Effects of oxic conditions on N₂O fluxes in forest soils (Heinävesi)

The process of coupled nitrification denitrification seems to be the dominant process for increasing N₂O fluxes (Butterbach-Bahl *et al.*, 2013) in organic soils whereby there is production of nitrate by nitrite oxidizers, which is immediately denitrified *in situ* by denitrifiers. Although denitrification is thought to be strictly anaerobic, it has been also observed to occur in well-aerated soils (Nielsen and MacDonald, 2013). Oxic treatment could have resulted in soil respiration which then could have led to anaerobic conditions in soil aggregates within the organic soil (Werner *et al.*, 2007). In aerobic soil aggregates, ammonium is oxidized to nitrate which then provides the substrate required for denitrifiers to reduce nitrate in the anaerobic soil aggregates (Kremen *et al.*, 2005). The occurrence of this process seemed to be supported by the generally declining trend in ammonium concentrations and consequent increase in nitrate concentrations and N₂O fluxes.

The process of aerobic denitrification is also known to play a pertinent role in environments not limited by oxygen (Patureau *et al.*, 2000) and can result in potential to produce N₂O (Lloyd, 1993). This process seems to be more prevalent in the mineral soils whereby nitrate seems to be consistent with steady concentrations of N₂O. Denitrifiers are facultative anaerobes and are also able to utilize oxygen when available (Hallin *et al.*, 2009) because aerobic respiration is more thermodynamically favourable. However, in aerobic conditions denitrification does not result in the final reduction of N₂O to N₂. N₂OR enzyme encoded by the NosZ gene in denitrifiers has the ability to reduce N₂O to N₂ (Sanford *et al.*, 2012) and when N₂OR enzyme is exposed to oxygen, the N₂OR enzyme becomes deactivated within a cell (Baggs *et al.*, 2009) resulting in N₂O not being reduced to N₂.

The net N₂O production potential was extremely high in organic soils compared to mineral soils which had nearly negligible net N₂O production. Organic soils could be richer in carbon substrates compared to the mineral soils which lead to comparatively higher net N₂O emissions in oxic conditions (Klemetsson *et al.*, 2005; Alm *et al.*, 2007). Moreover, the comparatively higher availability of nitrogen substrates and possibly higher degradable

carbon availability in organic soils than mineral soils could have resulted in the higher net N₂O production in aerobic conditions (Papen and Butterbach-Bahl, 1999).

5.3 Effects of Anoxic conditions on N₂O fluxes in peat (Salmisuo)

According to earlier studies, wetlands have been regarded as sources of N₂O (Davidson, 1991). However, recent studies including the results from our study, have shown that acidic wetlands have also the potential to act as a sink for N₂O (Inubushi *et al.*, 2003; Hadi *et al.*, 2005; Chapuis-Lardy *et al.*, 2007; Goldberg *et al.*, 2010).

In anoxic conditions, N₂O seemed to be produced and consumed by microbial denitrification in both peat layers but at different rates. This suggestion is supported by the physico-chemical properties of the peat soil including the high soil moisture content in the peat layers, which favours rapid denitrification, and the extremely low nitrate concentrations throughout the experiment, indicative of low nitrification potentials in both peat layers. Low nitrate concentrations have also been observed in earlier studies and associated with low rates of N₂O emissions (Martin and Holding 1978; Rosswall and Granhall 1980; Verhoeven 1986; Rangeley & Knowles 1988). Moreover, peat layers are sources of organic carbon, and dissolved organic carbon in anaerobic conditions can support the reduction of N₂O because denitrifying bacteria obtain their energy from soluble carbon needed for denitrification (International Plant Nutrition Institute, 2016).

The lower (10-20cm) layer of peat resulted in a quicker net consumption of N₂O than the upper (0-10cm) layer in the first 48 hours. This suggests that the denitrification potential and the potential to consume N₂O were higher in the lower (10-20cm) peat layer. This could be due to the soil characteristics and resulting microbial community differences in the two distinct layers. Peat decomposition is primarily driven by microorganisms and subsequent soil microbial community composition plays a key role in ecosystem processes and nutrient cycling (Balsler and Firestone, 2005; Bardget *et al.*, 2008; McGuire and Treseder, 2010). Differences in microbial communities have been noted to exist between peatland types (i.e. Bogs and Fens; Jaatinen *et al.*, 2007, Ausec *et al.*, 2009, Robroek *et al.*, 2014) and also among depths down the peat profiles (Morales *et al.*, 2006 and Jaatinen *et al.*, 2007). Microbial bacteria could be more predominant in the lower (10-20cm) peat layer than fungi and have been known to pre-dominate deeper more anoxic peat layers because they are able to utilize alternative electron acceptors (Killham and Prosser, 2007). This possible difference in

microbial community in the peat depths could have resulted in the different N₂O production/consumption rates.

The highly anoxic conditions of the peat soils combined with the anoxic treatment seemed to favour complete denitrification of N₂O to N₂ in the peat layers of Salmisuo soil (Klemedtsson *et al.*, 2005; Cuhel *et al.*, 2010; Braker and Conrad, 2011). Acid-tolerant denitrifiers in both peat layers likely reduced N₂O to N₂ (Palmer *et al.*, 2010, 2011) which resulted in a decrease in N₂O fluxes over time, but at different rates. Hence, it can be concluded that active denitrification-associated N₂O production and consumption occurs in acidic wetland (Goldberg *et al.*, 2008, 2010).

5.4 Effects of Oxic conditions on N₂O fluxes in peat (Salmisuo)

There were consistent N₂O fluxes emitted from both peat layers for the first 72 hours of the experiment. N₂O fluxes were generally higher in the lower (10-20cm) peat layer compared to the upper (0-10cm) peat layer. One reason could be that the oxic conditions could have resulted in ammonification and nitrification being dominant in the lower (10-20cm) peat layer resulting in N₂O fluxes due to inefficient denitrification (Williams and Wheatley, 1988; Regina *et al.*, 1996). This process seemed more dominant in the lower (10-20cm) peat layer compared to the upper (0-10cm) peat layer in the first 24 hours where both ammonium concentrations and nitrate concentrations were higher in the lower (10-20cm) peat layer than in the upper (0-10cm) peat layer. Throughout the experiment although ammonium concentrations fluctuated in the upper (0-10cm) peat layer, there was no discernable change in nitrate concentrations which were consistently low. The much higher ammonium concentrations in the upper (0-10cm) peat layer seem to have an inhibitory effect on the second step of nitrification whereby NO₂⁻ is oxidized to NO₃⁻ (Monaghan and Barraclough, 1992; Clough *et al.*, 2003; Ali *et al.*, 2013). Hence, this suggests that nitrification did not necessarily contribute to N₂O fluxes in the upper (0-10cm) peat layer and most N₂O fluxes in this layer probably occurred due to soil respiration.

The process of soil respiration by soil microbes in the peat layers could have contributed to CO₂ fluxes which in turn resulted in the consistent N₂O fluxes. In several studies, increased CO₂ fluxes have resulted in increased N₂O fluxes (Kammann *et al.*, 2008). Elevated CO₂ 1.7 times higher than ambient CO₂ has been noted to increase N₂O by 2-3 times (Arnone and Bohlen, 1998; Baggs *et al.*, 2003a, b) compared to ambient CO₂ conditions. This effect of subsequent increase in N₂O due to elevated CO₂ has also been observed to occur in soils

(Kettunen *et al.*, 2006). When CO₂ fluxes are increased, there could have been an increase in carbon allocation in the peat layers which provide energy for denitrifiers resulting in N₂O fluxes. Denitrification can then be also simultaneously occurring in anoxic microsites of both peat layers leading to N₂O consumption.

After 72 hours, denitrification seemed to be occurring in both peat layers resulting in a decrease in N₂O fluxes. As oxygen is consumed by the soil microbes in respiration, nitrate is then used in the lower (10-20cm) peat layer as the next favourable electron acceptor resulting in the decrease in N₂O fluxes. In the upper (0-10cm) peat layer, the low nitrate concentrations result in nitrate becoming limiting and with denitrifiers using N₂O as electron acceptors reducing N₂O to N₂ (Vanitchung *et al.*, 2011). Differences in microbial communities did not seem to play a key role in the N₂O fluxes in oxic treatment compared to anoxic treatment.

5.5 Effect of different electron donors on net N₂O consumption/potential in oxic conditions in mineral forest soil and peat

In our study, we also wanted to understand how the addition of different electron donors affected the net nitrous oxide consumption potential in the mineral forest soil and the lower (10-20cm) peat in oxic conditions. The mineral soil produced negligible N₂O in upland boreal forest while the 10-20cm had the most consumption potential amongst the 4 soil layers studied. Electron donors are pertinent because it has been shown that different electron donors stimulate distinct microorganisms (Ginige *et al.*, 2004; Osaka *et al.*, 2008; Baytshtok *et al.*, 2009). Therefore, it can be assumed that differences in microbial community structure and their tolerance/susceptibility to transient stressors could give rise to different emissions on different C-sources (Lu *et al.*, 2011). In soils, most denitrifying bacteria are heterotrophic and require carbon sources for cell growth and nitrate reduction (Lee and Rittmann, 2003). Carbon sources also have the ability to influence oxygen consumption rates in soils by directly increasing aerobic heterotrophic respiration or indirectly supporting the anaerobic production of reductants such as Fe(II) which then reacts with oxygen (Schlesinger, 2005).

The effect of electron donors in mineral forest soil and the lower (10-20cm) peat on net N₂O consumption potential in our study were variable. The microbial community at the lower (10-20cm) layer of peat seemed to have a preference for formate>succinate>propionate as electron donors in catalyzing N₂O reduction; with formate being most efficient in increasing net consumption of N₂O. The addition of these carbon sources seem to amplify the net reduction of N₂O 3-14 times higher compare to the oxic treatment with no addition of carbon

sources. This indicates that even a small addition of carbon sources have the ability to produce anaerobic conditions in soils that are partially or completely saturated (Schlesinger, 2005); enhancing complete denitrification and N₂O consumption. In the mineral soil, only propionate seemed to amplify N₂O consumption only by 1.5 times. This indicates that addition of propionate in the mineral soil layers could have resulted in anaerobic sites occurring within the soil aggregates which have been observed in upland forest soils (Højberg *et al.*, 1994, Schlesinger, 2005). However, in our study, the effect of supplemental electron donors on N₂O fluxes was unexpectedly not significant.

6. Conclusion

This study shows that denitrification plays a crucial role in regulating N₂O fluxes in boreal soils. The effect of oxygen on N₂O fluxes is different in boreal soil types and their layers. In anaerobic conditions, both forest and peat soils are able to consume N₂O although at different rates. In aerobic conditions, N₂O fluxes of forest and peat reacted differently with organic soils showing markedly increase in N₂O concentrations throughout the experiment and peat soils consuming N₂O after 3 days. Hence, these results indicate that N₂O fluxes in northern boreal soils are sensitive to aerobic/anaerobic conditions and are strongly affected by nitrate concentrations. The results from the addition of carbon sources under aerobic conditions to the mineral layer of the forest soil (negligible N₂O production) and to the lower peat layer (10-20cm) (high potential for N₂O consumption), implied that certain carbon sources can preferentially reduce N₂O fluxes.

However, it is difficult to elucidate exactly the processes that are occurring in the boreal soils. Additional stable isotope analysis and microbial community analysis is pivotal in determining exactly which processes and which microbes contributed to the N₂O fluxes. Moreover, N₂O fluxes can be markedly different depending on seasonal changes particularly in freeze-thaw cycles. It is important to note that this study was conducted primarily in the summer of 2014 and N₂O fluxes are generally known to be higher in the summers than during other seasons. Further studies of N₂O fluxes throughout the year and with these additional assessments could significantly improve our understanding of N₂O fluxes in northern boreal ecosystems.

7. References

- Aerts, R. and Ludwig, F. (1997). Water-table changes and nutritional status affect trace gas emissions from laboratory columns of peatland soils. *Soil Biology and Biochemistry*, 29 (11), 1691-1698.
- Ali, T. U., Kim, M. and Kim, D. J. (2013). Selective inhibition of ammonia oxidation and nitrite oxidation linked to N₂O emission with activated sludge and enriched nitrifiers. *J Microbiol Biotechnol* 5: 719-723
- Alm J., Shurpali, N. J., Minkkinen, K., Aro, L., Hytönen, J., Laurila, T., Lohila, A., Maljanen, M., Mäkiranta, P., Penttilä, T., Saarnio, S., Silvan, N., Tuittila, E-S. and Laine, J. (2007). Emission factors and their uncertainty for the exchange of CO₂, CH₄ and N₂O in Finnish managed peatlands. *Boreal Environ Res*, (12), 191–209
- Ambus, P., Zechmeister-Boltenstern, S., & Butterbach-Bahl, K. (2006) Sources of nitrous oxide emitted from European forest soils. *Biogeosciences*,(3)135–145
- Arnone III, J. A. and Bohlen, P. J. (1998). Stimulated N₂O flux from intact grassland monoliths after two growing seasons under elevated atmospheric CO₂. *Oecologia*, 116: 331-335
- Arp, D. J. and Stein, L. Y. (2003). Metabolism of inorganic N compounds by ammonia-oxidizing bacteria. *Crit Rev Biochem Mol Biol*. 38:471–495
- Aulakh, M. S., Doran, J. W., Walters, D. T. and Power, J. F. (1991). Legume residue and soil water effects on denitrification in soils of different textures. *Soil Biol. Biochem.* 23: 1161–1167
- Ausec, L., Kraigher, B. and Mandic-Mulec, I. (2009). Differences in the activity and bacterial community structure of drained grassland and forest peat soils. *Soil Biol. Biochem.* 41, 1874–1881
- Baggs, E. M. (2011). Soil microbial sources of nitrous oxide: recent advances in knowledge, emerging challenges and future direction. *Curr Opin Environ Sustainability*. 3:321–327
- Baggs, E. M., Ritcher, M., Cadisch, G. and Hartwig, U. A. (2003b). Denitrification in grass swards is increased under elevated atmospheric CO₂, *Soil Biol. Biochem.* 35: 729.732
- Baggs, E. M., Ritcher, M., Hartwig, U. A. and Cadisch, G. (2003a). Nitrous oxide emissions from grass swards during the eight years of elevated atmospheric pCO₂ (Swiss FACE), *Global Change Biol.*, 9: 1214-1222
- Baggs, L., Smales, L. C. and Bateman, J. E. (2010). Changing pH shifts the microbial sources well as the magnitude of N₂O emission from soil. *Biol Fertil Soils*. (46): 793.

- Balser, T. C. and Firestone, M. K. (2005). Linking microbial community composition and soil processes in a California annual grassland and mixed-conifer forest. *Biogeochemistry* 73: 395-415
- Bardgett, D. R., Freeman, C. and Ostle, J. N. (2008). Microbial contributions to climate change through carbon cycle feedbacks. *ISME* 2: 805-814
- Baumann, B. Snozzi, M., Zehnder, A. J. B. and VanderMeer, J. R. (1996). Dynamics of denitrification activity of *Paracoccus denitrificans* in continuous culture during aerobic-anaerobic changes. *J. Bacteriol* 178: 4367-4374
- Baytshtok, V., Lu, H., Park, H., Kim, S., Yu, R. and Chandran, K. (2009). Impact of varying electron donors on the molecular microbial ecology and biokinetics of methyltrophic denitrifying bacteria. *Biotechnol. Bioeng.* 102(6): 1527-36
- Bengtsson, G., and Bergwall, C. (2000). Fate of ¹⁵N labelled nitrate and ammonium in a fertilized forest soil. *Soil Biology & Biochemistry* 32:545-557.
- Berner, L. T., Alexander, H. D., Loranty, M. M., Ganzlin, P., Mack, M. C., Davydov, S. P., & Goetz, S. J. Biomass allometry for alder, dwarf birch, and willow in boreal forest and tundra ecosystems of far northeastern Siberia and north-central Alaska. *Forest Ecology and Management*, 337, 110–118.
- Blicher-Mathiesen, G. and Hoffmann, C. C. (1999). Denitrification as a sink for dissolved nitrous oxide in a freshwater riparian fen. *Journal of Environmental Quality*, (28), 257-262.
- Bobbink, R., Hornung, M. and Roelofs, J.G.M. (1998). The effects of air-borne nitrogen pollutants on species diversity in natural and semi-natural European vegetation. *Journal of Ecology*, 86, 717-738
- Bonin, P. and Gilewicz, M. (1991). A direct demonstration of ‘co-respiration’ of oxygen and nitrogen oxides by *Pseudomonas nautica*: some spectral and kinetic properties of the respiratory components. *FEMS Microbiol Lett* 80: 183-188
- Bouwman A.F., Boumans L.J.M. and Batjes N.H. (2002). Emissions of N₂O and NO from fertilized fields. Summary of available measurement data. *Global Biogeochemical Cycles*, 16(4): 1058 doi: 10.1029/2001GB001811.
- Braker, G. and Conrad, R. (2011). “Diversity, structure, and size of N₂O-producing microbial communities in soils – what matters for their functioning?” in *Advances in Applied Microbiology*, Vol. 75, eds Laskin A. I., Sariaslani S., Gadd G. M., editors. (San Diego: Elsevier Academic Press Inc.), 33–70
- Bremner, J. M., Breitenbeck, G. A. and Blackmer, A. M. (1981). Effect of anhydrous ammonia fertilizer on emission of nitrous oxide from soils. *J. Environ. Qual.* 10: 77–80
- Brierley, E.D.R., and M. Wood. (2001). Heterotrophic nitrification in an acid forest soil: isolation and characterisation of a nitrifying bacterium. *Soil Biology & Biochemistry* 33:1403-1409.

- Brumme, R., Verchot, L. V., Martikainen, P. J. and Potter, C. S. (2005). Contribution of trace gases nitrous oxide (N₂O), and methane (CH₄) to the atmospheric warming balance of forest biomes. *SEB Exp Biol Ser* 293-317
- Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R. and Zechmeister-Boltenstern, S. (2013). Nitrous oxide emissions from soils: how well do we understand the processes and their controls. *Philosophical Transactions of the Royal Society of B* 368: 20130122
- Butterbach-Bahl, K., Gasche, R., Breuer, L. and Papen, H. (1997). Fluxes of NO and N₂O from temperate forest soils: impact of forest type, N deposition and of liming on the NO and N₂O emissions. *Nutrient Cycle Agroecosystem*, (48), 79–90.
- Chang, C., Cho, C. M. and Janzen, H. H. (1998). Nitrous oxide emission from long-term manured soils. *Soil Sci Soc Am J* 62: 677–682
- Chapuis-Lardy, L., Wrage, N., Metay, A., Chottes, J. L., and Bernouxs, M. (2007). Soils, a sink for N₂O? A review, *Global Change Biol.*, 13, 1–17
- Chorover, J. (2002). Andisols. p. 64-67. In R. Lal (ed.) *Encyclopedia of soil science*. M. Dekker, New York, USA
- Christensen, S., Simkins, S. and Tiedje, J.M. (1990). Spatial variation in denitrification. *Soil Science Society Ame J* 54:1608–13.
- Clough, T. J., Sherlock, R. R. and Kelliher, F. M. (2003). Can liming mitigate N₂O fluxes from a urine-amended soil? *Australian Journal of Soil Research* 41: 439-457
- Conrad, R. (2002). Microbiological and biochemical background of production and consumption of NO and N₂O in soil, in: *Trace Gas Exchange in Forest Ecosystems*, edited by: Gasche, R., Papen, H. and Rennenberg, H. Kluwer Academic Publishers, Dordrecht, Netherlands, pp. 3-33
- Conrad, R. and Dentener, F.J. (1999). The application of the compensation point concepts in scaling of fluxes. In A.F. Bouwman (Ed.) *Approaches to scaling of trace fluxes in ecosystems*. *Developments of atmospheric science* 24. Elsevier, Amsterdam, pp.203-216
- Crutzen, J.P. and Oppenheimer, M. (2008). Learning about ozone depletion. *Climate Change* 89:143-154
- Cuhel, J., Simek, M., Laughlin, R. J., Bru, D., Cheneby, D., Watson, C. J. and Philippot, L. (2010). Insights into the effect of soil pH on N₂O and N₂ emissions and denitrifier community size and activity. *Appl. Environ. Microbiol.* 76, 1870–1878
- Davidson, E. A. (1991). Fluxes of nitrous oxide and nitric oxide from terrestrial ecosystems in *Microbial Production and Consumption of Greenhouse Gases*, eds Rogers J. E., Whitman W. B., editors. (Washington, DC: American Society for Microbiology;), 219–23

Davidson, E. A., and Verchot, L. V. (2000). Testing the Hole-in-the-Pipe Model of nitric and nitrous oxide emissions from soils using the TRAGNET Database. *Global Biogeochemical Cycles*, 14 (4), 1035-1043.

Duxbury, J.M. and Mosier, A. R. (1993): Status and issues concerning agricultural emissions of greenhouse gases. In *Agricultural Dimensions of Global Climate Change*. Kaiser, H.M., T.E. Drennen,(eds.), St. Lucie Press, Delray Beach, FL, pp. 229-258.

Easterling, W.E., P.R. Crosson, N.J.Rosenberg, M.McKenny, L.A.Katz, and K.Lemon, 1993: Agricultural impacts of and responses to climate change in the Missouri-Iowa-Nebraska-Kansas (MINK) region, Washington, DC, pp.221-228.

Eichner, M. J. (1990). Nitrous oxide emissions from fertilized soils: summary of available data. *J. Environ. Qual.* 19: 272–280

Erich, M. S., Bererie, A., and Duxbury, J. M. (1984). Activities of denitrifying enzymes in freshly sampled soils. *Soil Sci* 38: 25-32

Firestone, M. K., Davidson, E. A., Andreae, M. O., & Schimel, D. S. (1989). Microbiological basis of NO and N₂O production and consumption in soil. Exchange of trace gases between terrestrial ecosystems and the atmosphere. 7-21.

Flechard, C. R., Neftel, A., Jocher, M., Ammann, C. and Fuhrer, J. (2005). Bi-directional soil/atmosphere N₂O exchange over two mown grassland systems with contrasting management practices. *Global Change Biology* 11:2114-2127

Forster, P., Ramaswamy, V., Artaxo, P., Berntsen, T., Betts, R., Fahey, D. W., Haywood, J., Lean, J., Lowe, D. C., Myhre, G., Nganga, J., Prinn, R., Raga, G., Schulz, M. and van Dorland, R. (2007): Changes in Atmospheric Constituents and in Radiative Forcing. In: *Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change* [Solomon, S., D. Qin, Manning, M., Chen, Z., Marquis, M., Averyt, A. B., Tignor, M. and Miller H.L. (eds.)]. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.

Freeman, C., Liska, G., Ostle, N. J., Lock, M. A., Reynolds, B., and Hudson, J. (1996). Microbial activity and enzymic decomposition processes following peatland water table drawdown. *Plant and Soil*, 180 (1), 121-127.

Gebauer, R.L.E. and Ehleringer, J.R. (2000). Water and Nitrogen uptake patterns following moisture pulses in a cold desert community. *Ecology* 81: 1415-24

Ginige, M. P., Hugenholtz, P., Daims, H., Wagner, M., Keller, J. and Blackhall, L. L. (2004). Use of stable-isotope probing, full-cycle rRNA analysis, and fluorescence in situ hybridization-microautoradiography to study a methanol-fed denitrifying microbial community. *Appl. Environ. Microbiol.* 70: 588-596

- Godoy, R., Paulino, L., Oyarzún, C.L. and Boeckx, P. (2003). Atmospheric N deposition in central and southern Chile. An overview. *Gayana Botanica* 60:47-53
- Goldberg, S. D., Knorr, K.-H. and Gebauer, G. (2008). N₂O concentration and isotope signature along profiles provide deeper insight into the fate of N₂O in soils. *Isotopes Environ. Health Stud.* 44, 377–391
- Goldberg, S. D., Knorr, K.-H., Blodau, C., Lischeid, G. and Gebauer G. (2010). Impact of altering the water table height of an acidic fen on N₂O and NO fluxes and soil concentrations. *Glob. Chang. Biol.* 16, 220–233
- Goreau, T. J., Kaplan, W. A., Wofsy, S. C., McElroy, M.B., Valois F.W. and Watson, S. W. (1980). Production of NO₂⁻ and N₂O by nitrifying bacteria at reduced concentrations of oxygen, *Appl. Environ. Microbiol.*, 40(3), 526–532
- Gorham, E. (1991). Northern Peatlands: Role in Carbon Cycle and probable responses to climatic warming. *Ecological Applications* 1:2, 182-195
- Granli, T. and Bøckman, O. C. (1994) Nitrous oxide from agriculture. *Norwegian Journal of Agricultural Sciences, Supplement No. 12*, 1-128.
- Guthrie, T. F. and Duxbury, J. M. (1978). Nitrogen mineralization and denitrification in organic soils. *Soil Sci. Soc. Am. J.* 42: 908-912
- Hadi, A., Inubushi, K., Furukawa, Y., Purnomo, E., Rasmadi, M. and Tsuruta, H. (2005). Greenhouse gas emissions from tropical peatlands of Kalimantan, Indonesia. *Nutr. Cycl. Agroecosys.* 71, 73–80
- Hallin, S., Jones, M. C., Schloter, M., Philippot, L. (2009). Relationship between N-cycling communities and ecosystem functioning in a 50-year-old fertilization experiment. *ISME journal* 3: 597-605
- Hayatsu, M., Tago, K., and Saito, M. (2008). Various players in the nitrogen cycle: diversity and functions of the microorganisms involved in nitrification and denitrification. *Soil Science and Plant Nutrition*, 54 (1), 33-45
- Haynes, H. B. N. (1975). The stream and its valley. *Verh Int Verein Limnology* 19:1-15
- Hofstra, N. and Bouwman, A.F. (2005). Denitrification in agricultural soils: summarizing published data and estimating global annual rates. *Nutrient cycling in Agroecosystems* 72:267-278.
- Højberg, O., Revsbech, N.P. and Tiedje, J.M. (1994). Denitrification in soil aggregates analyzed with microsensors for nitrous oxide and oxygen. *Soil Science Society Am J* 58:1691–98
- Huang, Y., Zou, J., Zheng, X., Wang, Y. and Xu, X. (2004). Nitrous oxide emissions as influenced by amendment of plant residues with different C:N ratios. *Soil Biol and Biochem* 36(6): 973-981

Huygens, D., Boeckx, P., Templer, P., Paulino, L., Van Cleemput, O., Oyarzún, C., Müller, C. and Godoy, R. (2008) Mechanisms for retention of bioavailable nitrogen in volcanic rainforest soils. *Nature Geoscience* 1:543- 548

International Plant Nutrition Institute (2016). Nitrogen Notes. [http://www.ipni.net/publication/nitrogen-en.nsf/0/668099AE825517CB85257DD600054B8C/\\$FILE/NitrogenNotes-EN-5.pdf](http://www.ipni.net/publication/nitrogen-en.nsf/0/668099AE825517CB85257DD600054B8C/$FILE/NitrogenNotes-EN-5.pdf) , last accessed on 23rd March 2017.

Inubushi, K., Furukawa, Y., Hadi, A., Purnomo, E. and Tsuruta, H. (2003). Seasonal changes of CO₂, CH₄ and N₂O fluxes in relation to land-use change in tropical peatlands located in coastal area of South Kalimantan. *Chemosphere* 52, 603–608

IPCC, 2013: Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change [Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, Nauels A, Xia Y, Bex V, Midgley PM (eds.)]. Cambridge University Press, Cambridge, UK and New York, NY, USA, 1535 pp.

Jaatinen, K., Laiho, R., Vuorenmaa, A., Del Castillo, U., Minkkinen, K., Pennanen, T., Penttilä, T. and Fritze, H. (2007). Responses of aerobic microbial communities and soil respiration to water-level drawdown in a northern boreal fen. *Environmental microbiology* 10: 339–353

Johnston, C.A., Bridgham, S.D. and Schubauer-Berigan, J.P. (2001). Nutrient dynamics in relation to geomorphology of riverine wetlands. *Soil Science Society Am J* 65:557–77.

Jones, C. M., Welsh, A., Throbäck, I. N., Dörsch, P., Bakken, L. R., and Hallin, S. (2011). Phenotypic and genotypic heterogeneity among closely related soil-borne N₂-and N₂O-producing *Bacillus* isolates harboring the *nosZ* gene. *FEMS microbiology ecology*, 76 (3), 541-552.

Joosten, H. and Clarke, D. (2002). Wire use of mires and peatlands – Background and Principles including a Framework for Decision-making pp: 24-32

Kammann, C., Müller, C., Grünhage, L. and Jäger, H.-J. (2008). Elevated CO₂ stimulates N₂O emissions in permanent grassland. *Soil Biology & Biochemistry* 40, 2194-2205

Kang, L., Han, X., Zhang, Z., and Sun, O. J. (2007). Grassland ecosystems in China: review of current knowledge and research advancement, *Philos. T. R. Soc. B*, 362, 997–1008

Kellman, L. and Kavanaugh, K. (2008). Nitrous oxide dynamics in managed northern forest soil profiles: is production offset by consumption? *Biogeochemistry*, 90 (2), 115-128

Kent, A. D. and Triplett, E. W. (2002). Microbial communities and their interactions in soil and rhizosphere ecosystems. *Annu Rev. Microbiol.* 56:211

Kesik, M., Ambus, P., Baritz, R., Bruggemann, N., Butterbach-Bahl, K., Damm, M., Duyzer, J., Horvath, L., Kiese, R., Kitzler, B., Leip, A., Li, C., Pihlatie, M., Pilegaard, K., Seufert, S.,

- Simpson, D., Skiba, U., Smiatek, G., Vesala, T., and Zechmeister-Boltenstern, S. (2005). Inventories of N₂O and NO emissions from European forest soils, *Biogeosciences*, 2, 353–375,
- Kettunen, R., Saamio, S., Martikainen, P. J., and Silvola, J. (2006). Increase of N₂O fluxes in agricultural peat and sandy soil under elevated CO₂ concentration: concomitant changes in soil moisture, groundwater table and biomass production of *Phleum pretense*, *Nutr. Cycl. Agroecosyst.*, 74, 175–189
- Khalil, M. A. K. (1999). Non-CO₂ greenhouse gases in the atmosphere. *Annu. Rev. Energy Environ.* 24, 645–661.
- Killham, K. and Prosser, J. I. (2007). “The prokaryotes” in *Soil Microbiology, Ecology, and Biochemistry*, 3rd Edn, ed. Paul E. A., editor. (Oxford: Academic Press;), 119–144
- Kim, D-G, Sagar, S. and Roudier, P. (2012). The effect of nitrification inhibitors on soil ammonia emissions in nitrogen managed soils: a meta-analysis *Nutr. Cycl. Agroecosyst.* 93: 51–64
- Klemetsson, L., Von Arnold, K., Weslien, P. and Gundersen, P (2005). Soil CN ratio as a scalar parameter to predict nitrous oxide emissions, *Glob. Change Biol*, (11): 1142-1147
- Körner, H. and Zumft, W. G. (1989). Expression of denitrification in enzymes in response to the dissolved oxygen level and respiratory substrate in continuous culture of *Pseudomonas stutzeri*. *Appl. Environ. Microbiol* 55(7): 1670-1676
- Kowalchuk, G. A. and Stephen, J. R. (2001). Ammonia-oxidizing bacteria: a model for molecular microbial ecology. *Annu Rev Microbiol.* 55:485–529
- Kremen, A., Bear, J., Shavit, U. and Shaviv, A. (2005). Model demonstrating the Potential for Coupled Nitrification Denitrification in soil aggregates. *Environ. Sci. Technol* 39: 4180-4188
- Kroeze, C., Bouwman, L., and Slomp, C.P. (2007). Sinks for nitrous oxide at the Earth’s surface. In Reay, D., Hewitt, C.N., Smith, K. and Grace, J. (Eds). *Greenhouse gas sinks. UK/US: CABI International*
- Lapitan, R. L., Wanninkhof, R. and Mosier, A. R. (1999). Methods for stable gas flux determination in aquatic and terrestrial systems. In: Bouwman, A. F. (ed.) *Approaches to Scaling of Trace Gas fluxes in Ecosystems*. Elsevier Science, Amsterdam, pp 31-66
- Larsen, J. A. (1980). *The Boreal Ecosystem*. New York: Academic Press. pp500
- Lassey, K. and Harvey, M. (2007). Nitrous oxide: the serious side of laughing gas. *Water and Atmosphere* 15(2)
- Lee, K. and Rittmann, B. E. (2003). Effects of pH and precipitation on autohydrogenotrophic denitrification using the hollow-fibre membrane-biofilm reactor. *Water Res* 37: 1551-1556

- Li, C., Aber, J., Stange, F., Butterbach-Bahl, K. and Papen, H. (2000). A process-oriented model of N₂O and NO emissions from forest soils: 1. Model development, *J. Geophys. Res.*, 105(D4), 4369–4384
- Liikanen, A. and Martikainen, P. J. (2003). Effect of ammonium and oxygen on methane and nitrous oxide fluxes across sediment-water interface in a eutrophic lake. *Chemosphere* 52, 1287-1293
- Lloyd, D. (1993). Aerobic denitrification in soils and sediments: from fallacies to facts. *Trends. Ecol. Evol.* 8: 352-356
- Lloyd, D., Boddy, L. and Davies, K. J. P. (1987). Persistence of bacterial denitrification capacity under aerobic conditions – the rule rather than the exception. *FEMS Microbiol Ecol* 45: 185-190
- Lu, H., Nurruzzama, F., Ravindhar, J. and Chandran, K. (2011). Alcohol dehydrogenase expression as a biomarker of denitrification activity in activated sludge using methanol and glycerol as electron donors. *Environ. Microbiol.* 13(11): 2930-2938
- Maljanen, M., Hytönen, J., and Martikainen, P. J. (2001). Fluxes of N₂O, CH₄ and CO₂ on afforested boreal agricultural soils. *Plant and soil*, 231(1), 113-121.
- Maljanen, M., Liikanen, A., Silvola, J., and Martikainen, P. J. (2003). Measuring N₂O emissions from organic soils by closed chamber or soil/snow N₂O gradient methods. *European Journal of Soil Science*, 54(3), 625-631.
- Maljanen, M., Martikkala, M., Koponen, H. T., Virkajärvi, P., and Martikainen, P. J. (2007). Fluxes of nitrous oxide and nitric oxide from experimental excreta patches in boreal agricultural soil. *Soil Biology and Biochemistry*, 39(4), 914-920.
- Maljanen, M., Shurpali, N., Hytönen, J., Mäkiranta, P., Aro, L., Potila, H., & Martikainen, P. J. (2012). Afforestation does not necessarily reduce nitrous oxide emissions from managed boreal peat soils. *Biogeochemistry*, 108(1-3), 199-218
- Martikainen, P. J., Nykänen, H., Crill, P. and Silvola, J. (1993). Effect of a lowered water table on nitrous oxide fluxes from northern peatlands. *Nature*, 366(6450), 51-53.
- Martin, N. J., and Holding, A. J. (1978). Nutrient availability and other factors limiting microbial activity in the blanket peat. In: Heal, O. W. and Perkins, D. F. (eds.) *Production Ecology of British Moors and Montane Grasslands*. pp 113-135, Springer-Verlag, Berlin
- Matson, P. A., and Vitousek, P. M. (1987). Cross-system comparisons of soil nitrogen transformations and nitrous oxide flux in tropical forest ecosystems. *Global Biogeochemical Cycles*, 1(2), 163-170.
- McClain, E. M., Boyer, W. E., Dent, L. C., Gergel, E. S., Grimm, B. N., Groffmann, M. P., Hart, C.S., Harvey, W. J., Johnston, A. C., Mayorga, E., McDowell, H. W. and Pinay, G. (2013). Biogeochemical Hotspots and Hot moments at the Interface of Terrestrial and Aquatic Ecosystems. *Ecosystems* 6: 301-312

- McGuire, K. L. and Treseder, K. K. (2010). Microbial communities and their relevance for ecosystem models: decomposition as a case study. *Soil Biol. Biochem* 42: 529-535
- METLA. Forest Finland in Brief (2013). Forest statistical information, Vantaa: Finnish Forest Association
- Minami, K. and Fukushi, S. (1984). Methods for measuring N₂O flux from water surface and N₂O dissolved in water from agricultural land. *Soil Science and Plant Nutrition* 30:4 495-502
- Miyawaki, A. and Tüxen, R. (1977). *Vegetation science and environmental protection*. Maruzen, Tokyo
- Monaghan, R. and Barraclough, D. (1992). Some chemical and physical factors affecting the rate and dynamics of nitrification in urine-affected soil. *Plant and Soil*, 143(1), 11-18. Retrieved from <http://www.jstor.org/stable/42938048>, last accessed 22nd march 2017
- Morales, S. E., Mouser P. J., Ward N., Hudman, S. P., Gotelli, N. J., Ross, D. S. and Lewis, T. A. (2006). Comparison of bacterial communities in New England *Sphagnum* bogs using terminal restriction fragment length polymorphism (T-RFLP). *Microb. Ecol.* 52, 34–44
- Mosier, A., Kroeze, C., Nevison, C., Oenema, O., Seitzinger, S., and Van Cleemput, O. (1998). Closing the global N₂O budget: nitrous oxide emissions through the agricultural nitrogen cycle. *Nutrient cycling in Agroecosystems*, 52(2-3), 225-248.
- Mu Z, Huang A, Ni J, Xie D (2014) Linking Annual N₂O Emission in Organic Soils to Mineral Nitrogen Input as Estimated by Heterotrophic Respiration and Soil C/N Ratio. *PLoS ONE* 9(5): e96572. doi:10.1371/journal.pone.0096572
- Muñoz, C., Paulino, L., Monreal, C. and Zagal, E. (2010). Greenhouse gas (CO₂ and N₂O) emissions from soils: A review. *Chilean Journal of Agricultural Research* 70 N°3.
- Näsholm, T., Ekbald, A., Nordin, A., Giesler, R., Hogberg, M. and Hogberg, P. (1998). Boreal forest plants take up organic nitrogen. *Nature*, Vol. 392:914-916
- Németh, T., Abd El-Galil, A., Radimsky, L. and Baczó, G. Y. (1996). Effect of plant residues on ammonium and nitrate content of soils during incubation. In: van Cleemput, O., Hofman, G. and Vermoesen, A. (ed.) *Progress in Nitrogen Cycling Studies*, pp. 109–114, Springer, New York.
- Osaka, T., Shirotani, K., Yoshie, S. and Tsuneda, S. (2008). Effects of carbon source on denitrification efficiency and microbial community structure in a saline wastewater treatment process. *Water Res* 42: 3709-3718
- Palm, C. A., Alegre, J. C., Arevalo, L., Mutuo, P. K., Mosier, A. R., and Coe, R. (2002). Nitrous oxide and methane fluxes in six different land use systems in the Peruvian Amazon. *Global Biogeochemical Cycles*, 16 (4), 21-1.
- Palmer, K., Biasi, C. and Horn, M. A. (2011). Contrasting denitrifier communities relate to contrasting N₂O emission patterns from acidic peat soils in arctic tundra. *ISME J*.

- Palmer, K., Drake, H. L. and Horn, M. A. (2010). Association of novel and highly diverse acid-tolerant denitrifiers with N₂O fluxes of an acidic fen. *Appl. Environ. Microbiol.* 76, 1125–1134
- Papen, H. and Butterbach-Bahl, K. (1999). A 3-year continuous record of nitrogen trace gas fluxes from untreated and limed soil of an N-saturated spruce and beech forest ecosystem in Germany: 1. N₂O emissions. *Journal of Geophysical Research: Atmospheres* (1984–2012), 104(D15), 18487-18503.
- Parashar, D. C., Rai, J., Gupta, P. K., and Singh, N. (1991). Parameters affecting methane emission from paddy fields, *Indian J. Radio Space Phys.*, 20, 12–17.
- Parfitt, R.L., Scott, N.A., Ross, D.J., Salt, G.J., and Tate, K.R. (2003). Land-use change effects on soil C and N transformations in soils of high N status: comparisons under indigenous forest, pasture and pine plantation. *Biogeochemistry* 66:203-221.
- Parkin, T. B. (1987). Soil microsites as a source of denitrification variability. *Soil Science Society Am J* 51:1194-9
- Parkin, T.B. and Berry, E.C. (1994). Nitrogen transformations associated with earthworm casts. *Soil Biol Biochem* 26:1233–8.
- Patureau, D., Zumstein, E., Delgenes, P. J. and Moletta, R., (2000). Aerobic Denitrifiers Isolated from Diverse Natural and Managed Ecosystems. *Microb Ecol* (2000) 39:145–152
- Petersen, S. O., Nielsen, T. H., Frostegard, A. and Olesen, T. (1996). O₂ uptake, C metabolism and denitrification associated with manure hot-spots. *Soil Biol. Biochem.* 28: 341–349
- Philippot, L., Hallin, S., and Schloter, M. (2007). Ecology of denitrifying prokaryotes in agricultural soil. *Advances in Agronomy*, 96, 249-305
- Poth, M. and Focht, D. D. (1985). ¹⁵N kinetic analysis of N₂O production by a *Nitrosomonas europaea*. An examination of nitrifier denitrification. *Appl. Environ. Microbiol.* 49, 1134-1141
- Rangeley, A. and Knowles, R. (1988). Nitrogen transformation in a Scottish peat soil under laboratory conditions. *Soil. Biol. Biochem.* 20:385
- Ravishankara, A. R., Daniel, J. S. and Portmann, R. W. (2009). Nitrous oxide (N₂O): the dominant ozone-depleting substance emitted in the 21st century. *Science* 326: 123-125
- Regina, K., Nykänen, H., Silvola, J. and Martikainen, P. J. (1996). Fluxes of nitrous oxide from boreal peatlands as affected by peatland type, water table level and nitrification capacity. *Biogeochemistry* 35: 401-418 doi:10.1007/BF02183033
- Regina, K., Syväsalo, E., Hannukkala, A., Esala, M., (2004). Fluxes of N₂O from farmed peat soils in Finland. *Eur. J. Soil Sci.* 55, 591–599.

- Richardson, D., Felgate, H., Watmough, N., Thomson, A., and Baggs, E. (2009). Mitigating release of the potent greenhouse gas N₂O from the nitrogen cycle—could enzymic regulation hold the key? *Trends in biotechnology*, 27(7), 388-397
- Robertson, G. P. (1989). Nitrification and denitrification in humid tropical ecosystems: potential controls on nitrogen retention. In: Proctor, J. (eds.) *Mineral nutrients in tropical forest and savanna ecosystems*. Blackwell Scientific, Cambridge, Massachusetts, USA.
- Robertson, G.P. and Tiedje, J.M. (1984). Denitrification and nitrous oxide production in successional and old-growth Michigan forests. *Soil Science Society of America Journal* 48:383-389
- Robertson, L. A. and Kuenen, J. G. (1984). Aerobic denitrification – a controversy revived. *Arch. Microbiol.* 139: 351-354
- Robroek, M. J. B., Wubs, J. R. E., Martí, M., Zajac, K., Andersen, P. J., Andersson, A., Börjesson, G., Bragazza, L., Dise, B. N., Keuskamp, A. J., Larsson, M., Lindgren, P.-E., Mattiasson, P., Solomonsson, J., Sundberg, C., Svensson, H. B. and Verhoeven, A. T. J. (2014). Microclimatological consequences for plant and microbial composition in *Spaghnum*-dominated peatlands. *Boreal Env. Research* 19: 195-208
- Rosenkranz, P., Brüggemann, N., Papen, H., Xu, Z., Seufert, G., and Butterbach-Bahl, K. (2006). N₂O, NO and CH₄ exchange, and microbial N turnover over a Mediterranean pine forest soil. *Biogeosciences*, 3(2), 121-133.
- Rosswall, T., and Granhall, U. (1980). Nitrogen cycling in a subarctic ombrotrophic mire. *Ecol. Bull. (Stockholm)* (30): 209-234
- Rumney, G.R. (1968). *Climatology and the World's Climates*. New York: MacMillan. pp656.
- Ryden, J. C. (1981). N₂O exchange between a grassland soil and the atmosphere. *Nature*, (292), 235-365.
- Ryden, J. C. (1983). Denitrification loss from a grassland soil in the field receiving different rates of nitrogen as ammonium nitrate. *Journal of Soil Science* 34, 355-365.
- Rydin, H. and Jeglum, J. K. (2013). *The biology of peatlands*, 2e. Oxford: Oxford University Press.
- Sanford, R. A., Wagner, D. D., Wu, Q., Chee-Sanford, J. C., Thomas, S. H., Cruz-Garcia, C., Rodriguez, G., Massol-Deyá, A., Krishnani, K. K., Ritalahti, M. K., Nissen, S., Konstantinidis, T. K. and Löffler, F. E. (2012). Unexpected nondenitrifier nitrous oxide reductase gene diversity and abundance in soils. *Proc Natl Acad Sci. USA* 109(48): 19709-19714
- Savage, K., Phillips, R., and Davidson, E. (2014). High temporal frequency measurements of greenhouse gas emissions from soils, *Biogeosciences*, 11, 2709-2720

- Schlesinger, H. W. (2005). *Biogeochemistry. Treatise on Geochemistry: Volume 8*. Elsevier, Amsterdam
- Schlesinger, W. H. (2013). An estimate of the global sink for nitrous oxide in soils. *Global Change Biology*, 19 (10), 2929-2931
- Schurgers, G., Mikolajewicz, U., Groger, M., Maier-Reimer, E., Vizcaino, M. and Winguth, A. (2006). Dynamics of the terrestrial biosphere, climate and atmosphere CO₂ concentration during interglacials: a comparison between Eemian and Holocene. *Climate of the Past* 2:205-220
- Seech, A.G. and Beauchamp, E.G. (1988). Denitrification in soil aggregates of different sizes. *Soil Science Society Am J* 52:1616–21.
- Seitzinger, S. P. (1988). Denitrification in freshwater and coastal marine ecosystems: ecological and geochemical significance. *Limnol Oceanogr.* 33: 702-724
- Shapleigh, J. P. (2006). The denitrifying prokaryotes. In Dworkin, M. Falkow, S. Rosenberg, F. Schliefer, K. H., eds. *The prokaryotes. Vol. 2: Ecophysiology and Biogeochemistry*. New York: Springer, pp769-792
- Simek, M. and Cooper, J. E. (2002). The influence of soil pH on denitrification: progress towards the understanding of this interaction over the last 50 years. *Eur. J. Soil Sci.* 53, 345–354
- Skiba, U. and Ball, B. (2002). The effect of soil texture and soil drainage on emissions of nitric oxide and nitrous oxide. *Soil Use. Manag.* 18, 56-60
- Skiba, U., Sheppard, L., Pitcairn, C. E. R., Leith, I., Crossley, A., van Dijk, S., Kennedy, V. H. and Fowler, D. (1998). Soil nitrous oxide and nitric oxide emissions as indicators of elevated atmospheric N deposition rates in seminatural ecosystems. *Environ. Poll.* 102: 457-461
- Sommerfeld, R. A., Mosier, A. R., and Musselman, R. C. (1993). CO₂, CH₄ and N₂O flux through a Wyoming snow pack and implications for global budgets.
- Svensson, B.H., Bostrom, U. and Klemetsson, L. (1986). Potential for higher rates of denitrification in earthworm casts than in the surrounding soil. *Biol Fertil Soils* 2:147–9.
- Syakila, A. and Kroeze, C. (2011). The global nitrous oxide budget revisited, *Greenhouse Gas Measurement and Management*, 1:1, 17-26
- Teepe, R., Brumme, R., and Beese, F. (2000). Nitrous oxide emissions from frozen soils under agricultural, fallow and forest land. *Soil Biology and BioChemistry*, 32(11), 1807-1810.
- Ullah, S., Fraiser, R., King, L., Picotte-Anderson, N. And Moore, T.R. (2008). Potential fluxes of N₂O and CH₄ from soils of three forest types in Eastern Canada. *Soil Biology and BioChemistry*, 40(4), 986-994

- Vanitchung, S., Conrad, R., Harvey, N. W. and Chidthaisong, A. (2011) Fluxes and production pathways of nitrous oxide in different types of tropical forest soils in Thailand. *Soil Science and Plant Nutrition*, 57 (5), 650-658.
- Velthof, G. L., Brader, A. B. and Oenema, O. (1996). *Plant Soil* 181: 263. doi:10.1007/BF00012061
- Verchot, L. V., Davidson, E. A., Cattânio, H., Ackerman, I. L., Erickson, H. E., and Keller, M. (1999). Land use change and biogeochemical controls of nitrogen oxide emissions from soils in eastern Amazonia. *Global Biogeochemical Cycles*, 13(1), 31-46.
- Verhoeven, J. T. A. (1986). Nutrient dynamics in minerotrophic peat mires. *Aquat. Bot.* (25): 117-137
- Vitousek, P. M., Aber, J. D., Howarth, R. W., Likens, G. E., Matson, P. A., Schindler, D. W., Schlesinger, W. H., and Tilman, D. G (1997). Human alteration of the global nitrogen cycle: sources and consequences, *Ecol. Appl.*, 7, 737.
- Vitt, D. H., Halsey, L. A. and Nicholson, B. J. (2005). Chapter 6: The Mackenzie River Basin. In Fraser, L. H. and Keddy, P. A. (Eds.), *The World's Largest Wetlands: Ecology and Conservation* 166:198. Cambridge University Press
- von Arnold K, Nilsson, M., Hanell, B., Weslien, P. and Klemedtsson, L. (2005). Fluxes of CO₂, CH₄ and N₂O from deciduous forests on organic soils. *Soil Biology and Biochemistry*, (37), 1059–1071
- Voss, M., Bange, W. H., Dippner, W. J., Middelburg, J. J., Montoya, P. J. and Ward, B (2013). The marine nitrogen cycle: recent discoveries, uncertainties and the potential relevance of climate change. *Phil. Trans. R. Soc.*
- Ward, B. B. (2013). Update of Nitrification in *Encyclopedia of Ecology*. 2511-2518. https://www.princeton.edu/nitrogen/publications/pdfs/Ward_2015_Nitrification.pdf last accessed 23rd march 2017
- Werner, C., Kiese, R. and Butterbach-Bahl, K. (2007). Soil-atmosphere exchange of N₂O, CH₄, and CO₂ and controlling environmental factors for tropical rain forest sites in western Kenya. *Journal of Geophysical Research: Atmospheres* (1984–2012), 112(D3).
- Williams, B. L. and Wheatley, R. E. (1988). Nitrogen mineralization and water-table height in oligotrophic deep peat. *Biol. Fertil. Soils* 6:141
- Williams, E. J., Hutchinson, G. I. and Fehsenfeld, F. C. (1992). NO_x and N₂O emissions from soil. *Global Biogeochemical Cycles* 6: 351-388
- Wolf, B., Zheng, X., Brüggemann, N., Chen, W., Dannenmann, M., Han, X., Sutton, M. A., Wu, H., Yao, Z. and Butterbach-Bahl, K. (2010). Grazing-induced reduction of nitrous oxide release from continental steppe. *Nature* 464, 881-884

Wrage, N., G .L. Velthof, M. L. van Beusichem, and O. Oenema. (2001). Role of nitrifier denitrification in the production of nitrous oxide. *Soil Biology and Biochemistry* 33:1723-1732.

Zechmeister-Boltenstern, S., Hahn, M., Meger, S. and Jandl, R. (2002). Nitrous oxide emissions and nitrate leaching in relation to microbial biomass dynamics in a beech forest soil. *Soil Biol. Biochem.* 34(6): 823-832

Zhuang, Q., Lu, Y., and Chen, M. (2012). An inventory of global N₂O emissions from the soils of natural terrestrial ecosystems. *Atmospheric Environment*, (47), 66-75

Zumft, W. G. (1997). Cell biology and molecular basis of denitrification. *Microbiology and molecular biology reviews*, 61(4), 533-61