Open-Source Python Module for Automated Preprocessing of Near Infrared Spectroscopic Data

Jari Torniainen, Isaac O. Afara, Mithilesh Prakash, Jaakko K. Sarin, Lauri Stenroth, Juha Töyräs

PII: S0003-2670(20)30204-X

DOI: https://doi.org/10.1016/j.aca.2020.02.030

Reference: ACA 237462

To appear in: Analytica Chimica Acta

Received Date: 14 September 2019

Revised Date: 20 December 2019

Accepted Date: 12 February 2020

Please cite this article as: J. Torniainen, I.O. Afara, M. Prakash, J.K. Sarin, L. Stenroth, J. Töyräs, Open-Source Python Module for Automated Preprocessing of Near Infrared Spectroscopic Data, *Analytica Chimica Acta*, https://doi.org/10.1016/j.aca.2020.02.030.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

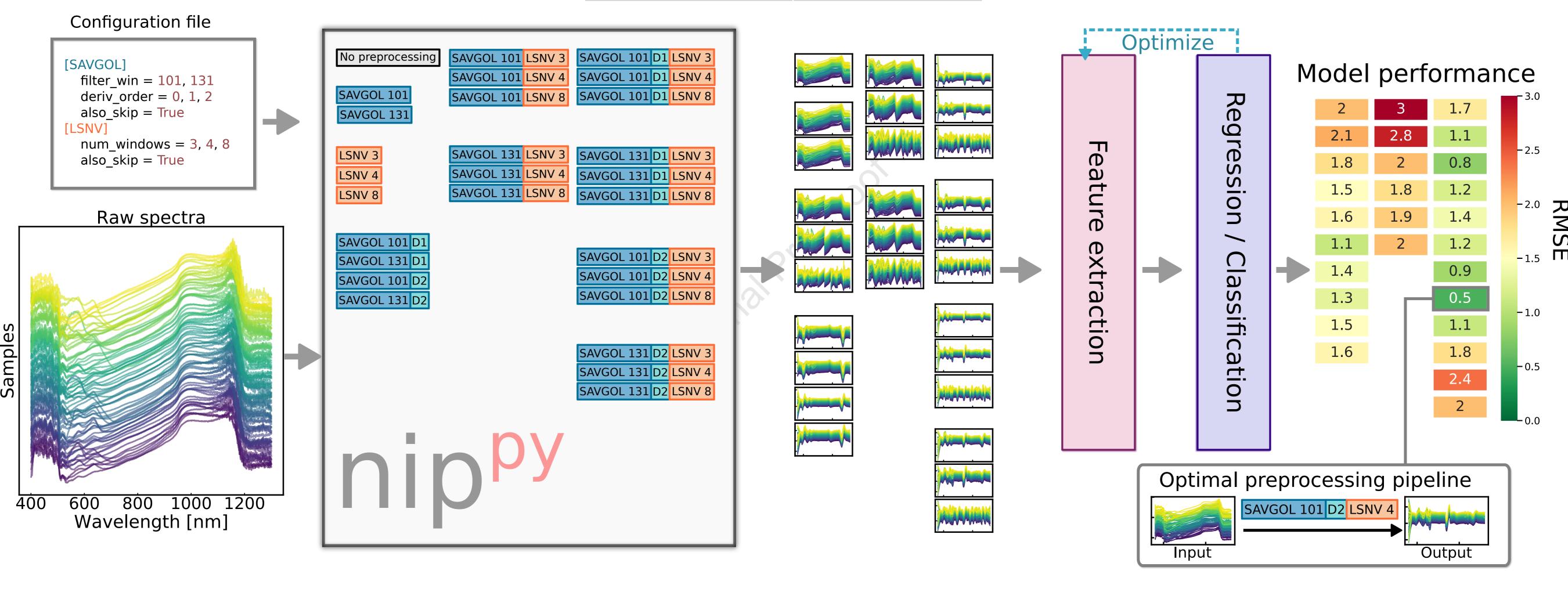
© 2020 Published by Elsevier B.V.



Credit Author Statement

Jari Torniainen: Investigation; Software; Writing - original draft Isaac O. Afara: Software; Writing - review & editing Mithilesh Prakash: Validation; Writing - review & editing Jaakko K. Sarin: Validation; Writing - review & editing Lauri Stenroth: Supervision; Writing - review & editing Juha Töyräs: Supervision; Project administration; Writing - review & editing

ounding





Open-Source Python Module for Automated Preprocessing of Near Infrared Spectroscopic Data

Jari Torniainen^{a,b}, Isaac O. Afara^{a,b}, Mithilesh Prakash^{a,b}, Jaakko K. Sarin^{a,b}, Lauri Stenroth^a, Juha Töyräs^{a,b,c}

^aDepartment of Applied Physics, University of Eastern Finland, Kuopio, Finland ^bDiagnostic Imaging Center, Kuopio University Hospital, Kuopio, Finland ^cSchool of Information Technology and Electrical Engineering, The University of Queensland, Brisbane, Australia

Abstract

Near infrared spectroscopy (NIRS) is an analytical technique for determining the chemical composition or structure of a given sample. For several decades, NIRS has been a frequently used analysis tool in agriculture, pharmacology, medicine, and petrochemistry. The popularity of NIRS is constantly growing as new application areas are discovered. Contrary to mid infrared spectral region, the absorption bands in near infrared spectral regions are often non-specific, broad, and overlapping. Analysis of NIR spectra requires multivariate methods which are highly subjective to noise arising from instrumentation, scattering effects, and measurement setup. NIRS measurements are also frequently performed outside of a laboratory which further contributes to the presence of noise. Therefore, preprocessing is a critical step in NIRS as it can vastly improve the performance of multivariate models. While extensive research regarding various preprocessing methods exists, selection of the best preprocessing method is often determined through trial-and-error. A more powerful approach for optimizing prepro-

Preprint submitted to Analytica Chimica Acta

February 15, 2020

cessing in NIRS models would be to automatically compare a large number of preprocessing techniques (e.g., through grid-search or hyperparameter tuning). To enable this, we present, nippy, an open-source Python module for semi-automatic comparison of NIRS preprocessing methods (available at https://github.com/uef-bbc/nippy). We provide here a brief overview of the capabilities of nippy and demonstrate the typical usage through two examples with public datasets.

Keywords: Near infrared spectroscopy, Preprocessing, Chemometrics

1 1. Introduction

Near infrared spectroscopy (NIRS) is a widely used vibrational spectro-2 scopic technique for quantitative evaluation of the composition and structure 3 of a given sample. In NIRS, the target sample is illuminated with near in-4 frared (NIR) light (750 – 2500 nm wavelength range) and the reflected and 5 backscattered light are measured with a spectrometer. NIR-active molecu-6 lar bonds in the sample absorb the incoming light at different overtone and combination spectral bands, thus producing the NIR absorbance spectrum. 8 Compared to other infrared spectroscopy methods, NIRS has increased pene-9 tration depth and less stringent requirements for sample preparation [1]. The 10 robustness and portability of NIRS devices enables their use as diagnostic 11 probes outside laboratory environments [1, 2]. NIRS is a relatively old an-12 alytical technique with pertinent research spanning well over three decades. 13 Today, NIRS is utilized heavily in multiple fields, including agriculture [3], 14 food processing [4], pharmaceutical industry [5], and medical research [6]. 15

¹⁶ The objective of a typical NIRS analysis is to relate the measured NIR

spectra to a target property of the sample which is either directly or indirectly 17 linked to its chemical composition or structure. For instance, a common ex-18 ample is the determination of the octane number of gasoline samples from the 19 corresponding NIR spectrum [7]. In this example, the number of CH-bonds in 20 hydrocarbons, like methyl and methylene, correlates with the gasoline octane 21 rating and can, therefore, be detected using NIRS. Compared to traditional 22 mid infrared spectroscopy, the NIR spectrum is more complex and difficult 23 to analyse due to the presence of overlapping overtones and combination 24 bands. The proper analysis of NIR spectral data, therefore, requires mul-25 tivariate statistical models. On the other hand, the robust-nature of NIRS 26 instrumentation enables integration of sensors for continuous monitoring of 27 chemical processes or as portable scanners for performing measurement and 28 analysis of solid samples on the field (e.g., soil or crops analysis). The best 29 practices in analysing NIRS data have been the topic of active research in 30 the field of chemometrics [1]. 31

The canonical analysis process of regression-based NIRS models is very 32 similar to other chemometric applications. First, a training set of data (i.e., 33 the calibration set) is collected by measuring both the NIR spectrum and 34 the desired target property. The calibration set should contain a relatively 35 large number of samples (e.g., N = 100 - 200, estimate derived from recom-36 mendations by Burns et al.[8] and ISO 12099:2017[9]) and cover the entire 37 current (and future) range of expected variation of the spectra and the target 38 property. The calibration dataset is used to construct a calibration model, 39 capable of predicting the target property of future samples. The resulting 40 calibration model should be reported with appropriate figures of merit (root 41

⁴² mean squared error, RMSE; coefficient of determination, R²; etc.) [10, 11]
⁴³ that approximate the real-world performance.

The workflow for generating the calibration model can be divided into 44 three main steps: preprocessing, variable selection (also known as frequency 45 selection or feature extraction), and calibration (i.e., training the classifi-46 cation or regression model). Preprocessing aims to remove all sources of 47 uninformative variance (e.g., instrumentation and scattering effects) from 48 the measured spectrum. Typical preprocessing steps include normalization, 49 linearization, and smoothing [12]. The variable selection aims to reduce the 50 full feature space (i.e., different wavelengths) to a subset of most important 51 features. Variable selection methods include various decomposition methods, 52 such as principal and independent component analysis, sequential methods 53 like uninformative variable elimination, and optimization techniques like ge-54 netic algorithms [13, 14]. In the last two cases, the process of variable selec-55 tion is performed in tandem with the calibration step. Finally, the calibration 56 step trains a classification/regression model which maps the extracted spec-57 tral features to explain the property of interest. Each of the three steps has 58 multiple viable methods and configurable parameters which can be tuned 59 for achieving the best possible predictive performance. The combination of 60 preprocessing, variable selection, and calibration producing the model with 61 the best performance depends heavily on the instrumentation and intended 62 application. 63

As the workflow for constructing the calibration model dictates how well the technique will perform, a large number of studies have focused on optimizing this process. A substantial amount of time and effort has been

used to compare different preprocessing [15, 16, 17, 12], feature extraction 67 [14, 13, 18], and classification methods [19, 20, 21] in order to build opti-68 mal predictive NIRS models. More recently, the effectiveness of different 69 combinations of preprocessing methods (often termed "preprocessing strate-70 gies") has been investigated. Engel et al. [22] produced the first critical review 71 of chemometric preprocessing strategies which demonstrated that the model 72 performance between different preprocessing strategies can vary substantially 73 and highlighted the drawbacks in methods currently in use for selecting these 74 strategies. A study by Gerretzen et al. [23] addressed the problem by utilizing 75 *design-of-experiments* approach for selecting the best preprocessing strategy. 76 This approach was later expanded to cover both preprocessing and variable 77 selection, with the aim of improving not only the model performance but also 78 its interpretability [24]. Earlier, Bocklitz et al. [25] presented a similar solution 79 based on genetic algorithms for finding the optimal preprocessing strategy in 80 Raman spectroscopy. Earlier investigations of various methods have resulted 81 in a collective consensus of good methods in the field of NIRS research. How-82 ever, in order to combine these methods to form the best possible calibration 83 model, each part of the model (i.e., preprocessing, variable selection, and 84 calibration) should be optimized individually for each application. 85

Recent advances in the field of machine learning have produced powerful methods (such as automated learning and hyperparameter tuning) for seeking further optimized models. The optimization of calibration models, however, has mainly focused on tuning the variable selection and classification/regression parts of the process with little focus on the preprocessing stage. While preprocessing has a critical impact on model outcomes, it is often not subjected to similar pruning. Likewise, the effect of different parameter values (e.g., filtering window length) on the model performance is
rarely exhaustively studied or their effects reported.

To help find an effective combination of preprocessing methods, we present 95 **nippy**: a toolbox to rapidly test different preprocessing combinations in con-96 junction with different machine learning methods. nippy is a Python 3.6+ 97 module, open source under the MIT license, and fully compatible with the 98 Python scientific stack (e.g., scikit-learn [26] machine learning module 99 and scipy [27]). While nippy is intended for spectral preprocessing, it can 100 be combined with the powerful outlier detection (e.g., elliptic envelopes, iso-101 lation forests, etc.), variable selection (e.g., variance thresholding, recursive 102 feature elimination, etc.), and model validation (e.g., classification/regression 103 metrics, dummy estimators, etc.) methods of the scikit-learn module in 104 order to produce state-of-the-art calibration models. It implements all rec-105 ommended preprocessing methods (such as scatter correction, smoothing, 106 and computing derivatives) as well as a framework for testing the effects of 107 different combinations of preprocessing methods and their parameters. To 108 be clear, we are not aiming to produce a comprehensive benchmarking of 109 different preprocessing methods but rather provide a tool for researchers and 110 practitioners to find an effective preprocessing strategy for their data. 111

We provide here a brief overview of the module and its capabilities and demonstrate the typical usage through two example cases using previously published datasets. Full documentation and the source code of nippy are also available at https://github.com/uef-bbc/nippy.

6

¹¹⁶ 2. Materials and methods

117 2.1. Overview of nippy

The near infrared preprocessing in Python toolbox (abbreviated as nippy) 118 is a Python module that enables the user to easily define multiple different 119 preprocessing strategies for NIRS data. The module is written in Python 120 3.6 and should, therefore, be compatible with all later versions. The module 121 is also fully compatible with the data format used by the scikit-learn-122 module, enabling easy integration with powerful machine learning methods. 123 A standard configuration file provided with the modules can be used to install 124 nippy automatically using the Python package management system (i.e., pip 125 or conda). 126

The nippy module consists of two main components: a preprocessing 127 module and a handler function. The preprocessing module collects all cur-128 rently accepted preprocessing methods under one file. Inputs to these meth-129 ods have been standardized and the general naming convention from NIRS 130 literature has been used when possible. To improve performance and reduce 131 the burden of maintenance, the best methods from existing larger modules 132 (such as sklearn and numpy) have been used. The modular structure of 133 preprocessing methods also enables the user to implement custom prepro-134 cessing methods if they so desire. An overview of the preprocessing methods 135 currently provided by **nippy** can be found in the section 2.2. The handler 136 component of **nippy** constructs preprocessing pipelines per users requests. 137 The user provides a list of methods and a list of parameters for each method 138 through a configuration file. The handler parses the configuration and pro-139 duces all possible permutations which can then be executed as a combination 140

of appropriate functions of the preprocessing module. The default usage of
the handler function is covered in section 2.3.

143 2.2. Preprocessing methods

In total, six different preprocessing categories are supported in nippy and
 they are performed in the following order.

146 1. Clipping

¹⁴⁷ 2. Scatter correction

¹⁴⁸ 3. Smoothing

¹⁴⁹ 4. Derivatives

150 5. Trimming

151 6. Resampling

It should be noted, that the order in which the preprocessing operations 152 are performed can have an effect on the model performance. The order se-153 lected for nippy represents the consensus among the authors and is based on 154 the order typically found in literature. If needed, the order of the operations 155 can be changed by modifying the source-code (details for this can be found in 156 the online documentation). Some of the above-mentioned operations (such 157 as scatter correction) can contain multiple alternative methods. The han-158 dler component of nippy will make sure that only one operation from each 159 category is included in each of the preprocessing pipes. 160

¹⁶¹ Clipping operation removes or substitutes data points with values ex-¹⁶² ceeding the user-defined threshold. This method is intended to eliminate ¹⁶³ short spike-like artifacts that might otherwise distort further analysis. As ¹⁶⁴ this method induces short discontinuities in the data, it is advisable to only use it for gross artifacts. Larger noise-contaminated regions can be removed
with the trimming operation.

Scatter correction methods aim to counter the particle size effect. In 167 nippy, the three standard methods (standard normal variate, SNV [28]; mul-168 tiplicative scatter correction, MSC [29]; and normalization [12]) have been 169 implemented. The non-parametric version of SNV, known as robust normal 170 variate (RNV) is also implemented [30]. In SNV, the correction is performed 171 according to the mean and standard deviation of the spectrum and in RNV 172 (which is intended for more noisy data) according to the median value and 173 user-specified inter-quartile interval. In addition, localized version of SNV 174 (LSNV) [31] is also included, where SNV operation is performed piece-wise 175 inside user-defined spectral windows. In MSC, correction is only possible to 176 the mean of the spectra. The extended version of MSC (EMSC) can also 177 be used, which takes into account both the linear and quadratic terms when 178 performing the correction. [32, 33] Normalization of the spectra can be per-179 formed to the value ranges specified by the user (e.g., between 0 and 1). If 180 no normalization range is provided, each spectrum is normalized with the re-181 spective Euclidean norm. Instead of scatter correction, nippy also supports 182 baseline correction which only mean-centers the spectra. 183

Smoothing of the NIR spectra can be helpful for removing environmental or instrumentation-related noise. nippy provides both convolution-based and Savitzky-Golay filtering [34] as smoothing methods. The Savitzky-Golay filter is also able to return smoothed derivatives of the original spectrum.

Sometimes using the entire available range of wavelengths is not desired and instead the analysis is constrained to specific optical windows [35]. Trimming operation enables the extraction of continuous and non-continuous
wavelength regions from the full spectral data.

As a final step, the spectra can be resampled to a new spectral resolution using the Fourier method. This step can be useful, for instance, when combining spectra acquired with multiple devices having different spectral resolutions.

196 2.3. Usage of *nippy*

The default format for data entry in nippy is numpy-matrices. The module requires one vector specifying the wavelengths and a matrix containing the spectra. The first dimension of the matrix must correspond to the wavelengths while the second dimension corresponds to samples.

Pipelines in **nippy** are defined using a configuration file. Structure of 201 the configuration file follows the INI format (https://en.wikipedia.org/ 202 wiki/INI_file). In the configuration file, preprocessing methods are en-203 tered as sections and parameters as key-value pairs. Most parameters (such 204 as the length of the filtering window) accept multiple variations of the pa-205 rameter value which can be separated with a comma. An up-to-date list 206 of preprocessing methods and related parameters can be found in the on-207 line documentation (https://github.com/UEF-BBC/nippy/blob/master/ 208 CONFIGURATION.md). Reading the configuration file will produce a list of 209 nested dictionaries where the dictionaries contain the argument-value pairs 210 of the preprocessing functions. 211

Once the NIR data and the pipeline configuration have been loaded, they can be passed to nippy. The handler module of nippy then automatically applies all combinations of preprocessing methods defined in the configuration file. The operation returns a list of copies of the original data, each modified according to the methods in the pipeline list. The preprocessed data is returned as numpy-arrays, meaning it can be fed directly in most machine learning methods. The module also supports iteration through a *Preprocessor*-class, which returns one preprocessed version of the original data at a time.

nippy also supports data export to other platforms (such as MATLAB
or R). The export formats currently supported are MAT, CSV, and Pickle.
An in-depth description of the data structure in exported files can be found
from the online documentation.

225 3. Results

The basic operation of nippy (fig. 1) is demonstrated through two realworld examples. In both cases, the construction of a calibration model is optimized by finding the best NIRS preprocessing pipeline through grid search. Source code for both examples is available at https://github.com/ uef-bbc/nippy-aca-2019.

231 3.1. Example 1: Classification of Ethiopian barley variants using NIR

The first example case is a classification task with a publicly available NIR dataset. The dataset (originally published by Kosmowski et al. [36]) consists of NIR measurements of 1200 samples of Ethiopian barley from 24 different barley cultivar variants. The objective of the example is to classify the barley cultivar variant (Ardu 1260 B, Bahati, Bekoji-1, etc.) of a given sample based on the NIR spectrum. This example represents a typical agricultural ²³⁸ application where data collected in a field with a portable device is utilized²³⁹ for quality control.

As in Kosmowski et al., the dataset was first split into training (70%) and 240 testing (30%) sets. The test set was stratified over the 24 classes resulting in 241 15 samples per class. Preprocessing of the data was performed using nippy. 242 Similar to the original analysis by Kosmowski et al., classification of the sam-243 ples was performed using support vector machines (SVMs). A nu-regularized 244 SVM with a polynomial kernel function was trained separately for each in-245 dividual preprocessing pipeline and the hyperparameters of each model were 246 optimized using a five-fold cross-validation. As the initial inspection of the 247 NIR spectra looked noise-free and relatively flat (fig. 2a), the preprocessing 248 techniques were restricted to Savitzky-Golay filtering and two scatter correc-249 tion methods (SNV and RNV). The parameter combinations used to build 250 these preprocessing pipelines are listed in table 1. 251

In total, 38 different preprocessing combinations (table 1) and one base-252 line model without preprocessing were tested (see fig. 2a for original data 253 and 2b for different preprocessing methods). The effect of each preprocess-254 ing method was quantified by comparing the accuracies of hold-out test and 255 training datasets over different pipelines (fig. 3). The best overall accuracy 256 (82.6% for training and 87.2% for test set) was obtained using a preprocessing 257 pipeline with SNV scatter correction and a first-order derivative Savitzky-258 Golay filtering (3rd order polynomial, 11-point window length). 259

The baseline model without any preprocessing yielded an accuracy of 75.0% for training and 80.3% for test set. The confusion matrices of the classification results were computed for the baseline model and the best per-

forming preprocessing method (fig. 4). The classification accuracy of the best performing preprocessing pipeline was identical to the best performing model reported by Kosmowski et al. [36].

3.2. Example 2: Regression model for predicting the instantaneous modulus of equine articular cartilage

The second example focuses on complex and noisy data, where NIR spec-268 tra were measured from the articular cartilage surface of equine fetlock joints 269 (fig. 5). Articular cartilage is a layer of viscoleastic connective tissue cover-270 ing the ends of articulating bones within a joint. Material properties of the 271 cartilage layer (such as instantaneous modulus) are an important indicator of 272 joint health. Ability to determine the material properties of cartilage during 273 an arthroscopic procedure could have substantial diagnostic significance in 274 identifying healthy and degraded regions of the joint. 275

The dataset was originally published by Sarin et al. [37] and optimal regression and variable selection methods for this data were subsequently investigated by Prakash et al. [38]. NIRS measurements were performed on 869 points from the proximal phalanx and the metacarpal bone of five horses. Instantaneous modulus at each measurement point was determined using a custom material testing device (for details see[37]). The dataset and an in-depth description of different variables can be found in [39].

The objective of this example is to construct a calibration model capable of predicting the instantaneous modulus of cartilage from the NIR spectrum. An earlier investigation of model selection and regression methods for this dataset [38] determined that the best prediction performance ($R^2 =$ 0.51, RMSEP = 2.46 MPa) was obtained with a five-component partial least squares regression. The preprocessing used in that analysis consisted of a third order Savitzky-Golay filtering (25 nm window size) and second order spectral derivation. Identical regression technique was used here and the effect of preprocessing was investigated using nippy. The model was validated with the same holdout test method (N=70, 9% of the dataset) used in the original study[38]. Baseline test set prediction performance without any kind of pretreatment of the spectra was $R^2 = 0.25$ and RMSEP = 3.06 MPa.

The pipelines (i.e., parameter combinations) used for the preprocessing 295 consisted of 3rd order Savitzky-Golay filtering with (12 window sizes, up 296 to the 2nd derivative) and convolution filtering (12 window sizes). Parti-297 cle size effects were compensated using MSC, SNV, LSNV (four different 298 window sizes), and RNV (four different interquartile ranges). Preprocessing 299 was performed on the full spectral range, as well as subsets of 700 - 900 nm 300 and 850 - 1050 nm. The absence of each aforementioned preprocessing step 301 was also investigated by leaving them out of the analysis. In total, nippy 302 was used to generate 1618 comparable pipelines (table 2) which were then 303 compared using PLSR. Number of components for each PLSR model was 304 determined using five-fold cross-validation. Best test set prediction perfor-305 mance in terms of \mathbb{R}^2 was obtained using the wavelength range of 700 - 950306 nm. RNV scatter correction (85% - 15%) interquartile range), and a Savitzky-307 Golay filtering with 73 nm window size (see figs. 6A and 6B). In comparison 308 to the baseline PLSR model, the preprocessing pipeline increased the coef-309 ficient of determination by approximately 38% ($R^2 = 0.63$, RMSEP = 2.15) 310 MPa, see fig. 6C). Preprocessing also improved the residuals of the model in 311 terms of magnitude, homoscedasticity, and normality (fig. 6D). Compared 312

to the performance reported earlier with this dataset, optimization of the preprocessing step provided a modest improvement of approximately 12% in terms of R² and 13% in terms of RMSEP.

Preprocessing	Parameter	Values	
operation			
SNV	<pre>snv_type:</pre>	snv, rnv	
	also_skip:	True	
SAVGOL	filter_win:	11,21,51,101	2
	deriv_order:	0, 1, 2	
	poly_order:	3	
	also_skip:	True	

Table 1: Configuration parameters for example 1.

316 4. Discussion

Preprocessing of NIR spectrum is a fundamental part of any NIRS application. An optimized preprocessing protocol can substantially improve the predictive capabilities of NIRS models. In this paper, we presented an open-source Python module for semi-automatic exploration and comparison of different preprocessing strategies. Ideally, the tools introduced in this paper should cut down development time when researching or building new NIR-based analytical applications.

While improving the prediction performance is the main use of nippy, it also enhances understanding why some preprocessing improves the result by revealing additional details about the underlying phenomena. By comparing

Preprocessing	Parameter	Values
operation		varies
MSC	also_skip:	True
SNV	also_skip:	C True
RNV	iqr:	75-25, 85-15, 65-35, 70-30
	also_skip:	True
LSNV	num_windows:	3, 5, 7, 9
	also_skip:	True
SAVGOL	filter_win:	11,25,41,55,71,85,101,115,131,145,161,175
	deriv_order:	0, 1, 2
	poly_order:	3
	also_skip:	True
SMOOTH	filter_win:	11,25,41,55,71,85,101,115,131,145,161,175
	also_skip:	True
TRIM	bins:	700 - 950, 850 - 1050
	also_skip:	True

Table 2: Configuration parameters for example 2.

which methods work and which do not can yield more insight into what kind of instrumentation is needed by the application. For example, the substantial increase in accuracy as a result of derivation in the Ethiopian barley classification (section 3.1) most likely indicates that the differentiating factors are minute spectral peaks and not the baseline level of the signal.

³³² Software tools for chemometric analysis of NIRS data have existed for a ³³³ long time and range from proprietary analysis solutions, such as The Un-

scrambler X (Camo Analytics, Oslo, Norway), OPUS (Bruker Corporation, 334 Billerica, MA, USA), or Pirouette (Infometrix Inc., Bothell, WA, USA), to 335 open-source libraries. While proprietary software can be useful in industrial 336 applications, research of new NIRS analysis techniques is typically conducted 337 with data science and programming oriented methods using tools, such as 338 R, MATLAB, or Python. The main benefits of open-source tools are trans-339 parency, customizability, easier access, and lower cost. We have, therefore, 340 limited our comparison of nippy to other comparable free or open-source 341 tools, more specifically, prospectr [40] and ParLeS [41] 342

The prospectr is one of the most popular R packages for analysing visible 343 and NIR spectroscopic data. The package implements a set of preprocessing 344 and sampling functions in the R language. Preprocessing methods included in 345 prospectr are largely similar to those found in nippy. The sampling meth-346 ods present different techniques for selecting training, testing, and validation 347 sets for constructing the calibration models (e.g., Kennard-Stone sampling, 348 DUPLEX sampling, etc.). Variable selection and calibration methods are 340 not included in the prospectr package. ParLes is a shareware software solu-350 tion for constructing NIRS calibration models. It implements the full chain 351 of operations ranging from preprocessing and variable selection to training 352 calibration models. Like prospectr, various sampling methods for training 353 the models are also provided. 354

In comparison, nippy shares more features with the prospectr package than ParLeS, in the sense that they are both libraries containing functions for preprocessing NIRS data. Main difference between the two is that the prospectr package does not enable rapid iteration of multiple comparable

preprocessing pipelines. ParLes is more of a stand-alone tool for data ex-359 ploration that enables the user to perform multiple different manipulations 360 and analysis on the given data. Again, however, the ability to rapidly and 361 programmatically test the effect of different preprocessing pipelines is miss-362 ing. Furthermore, integration to other analysis platforms (e.g. Python or 363 MATLAB) can not be done directly. As ParLeS is a shareware application, 364 extending the analysis capabilities of the tool is impossible without the help 365 of the original author. 366

Several earlier chemometric studies have also indicated that preprocess-367 ing has a substantial impact on the performance spectroscopic models and 368 depends on such factors as: the preprocessing operations, parameters used, 369 and the order of operations [42, 17, 25, 22, 23, 24]. In addition, Engel et al. 370 [22] pointed out that the sequential optimization of a preprocessing pipeline 371 might not work, as the synergy between different operations can be hard to 372 predict in advance. These studies have also suggested different approaches 373 for finding a suitable preprocessing strategy from multiple comparable alter-374 natives. For instance, Wold et al. [42] suggested utilizing orthogonal signal 375 correction, where preprocessing methods aim to remove linearly uncorrelated 376 spectral components with respect to the target property. Solution proposed 377 by Xu et al. [17] used Monte Carlo sampling for selecting the best prepro-378 cessing pipeline from multiple alternatives. Other approaches have suggested 379 genetic algorithms [25] or design of experiments approach [23] for selecting 380 the preprocessing strategy. Combination of preprocessing and variable se-381 lection under the same optimization process has also been investigated [24]. 382 While exhaustive grid-search never fails to find the optimal preprocessing 383

combination for a given dataset (provided that the grid of operations and related parameters is dense enough), the operation can be very time-consuming. The proposed hyperparameter tuning methods can converge faster and can enable larger search space for parameters. As a final step, generalization of the optimal solution outside the training data should be verified in order to avoid overfitting.

While the current implementation of **nippy** enables the user to rapidly sift 390 through various combinations of preprocessing operations, it does not provide 391 actual feedback on which of the combinations is the most effective. This 392 feature was intentionally left out as metaheuristics (also known as automated 393 machine learning or hyperparameter search), a relatively recent and very 394 active field of research, can deal with higher level optimization of machine 395 learning pipelines. Metaheuristics can be utilized to find the best solution 396 to a problem by individually tuning the preprocessing, feature extraction, 397 and classification/regression. Several powerful tools, ranging from bayesian 398 optimization [43] to genetic programming [44], already exist for solving this 390 problem. Instead of competing with existing tools, **nippy** was built with focus 400 on compatibility with modern metaheuristic tools. As machine learning tools 401 are nowadays predominantly written in Python and use numpy-based matrix 402 structures as its base, the same approach was adopted here. The end goal was 403 to combine decades worth of domain knowledge gained from various NIRS 404 publications and combine it with powerful machine learning frameworks. 405

In the future, nippy could potentially be extended to cover other spectroscopic techniques as well. Raman and mid infrared spectroscopy are very similar to NIRS and, thus, benefit from the same preprocessing operations. However, as NIRS is often used in industrial and agricultural applications,
the most crucial need for optimized preprocessing is with this technique.

411 5. Conclusions

To conclude, a lot is known about the different ways to eliminate scatter-412 ing effects and external noise from NIR spectra. Due to the great diversity of 413 different NIRS applications, the best preprocessing strategy is often depen-414 dent on the intended use. Finding the correct combination of preprocessing, 415 variable selection, and calibration has been the focus of much recent research. 416 To facilitate the optimization of preprocessing for NIRS models, we have de-417 veloped nippy, a tool that enables rapid iteration of different preprocessing 418 combinations. We feel that tools, such as nippy, are important to chemomet-419 rics because they provide researchers easy access to current state-of-art NIR 420 preprocessing and thus enable them to focus on optimizing models instead 421 of reinventing the wheel. 422

423 6. Acknowledgements

This research received funding from the following sources: SCITECO Doctoral Programme of University of Eastern Finland, Kuopio University Hospital (VTR projects 5041750, 5041744, 5041778, and 5203111, PY210 Clinical Neurophysiology), the Academy of Finland (projects 267551, 310466, and 315820), Orion Research Foundation sr, and Finnish foundation of technology promotion (project 8193).

430 7. Contributions

431 Jari Torniainen: Investigation; Software; Writing - original draft

432 Isaac O. Afara: Software; Writing - review & editing

433 Mithilesh Prakash: Validation; Writing - review & editing

434 Jaakko Sarin: Validation; Writing - review & editing

435 Lauri Stenroth: Supervision; Writing - review & editing

Juha Töyräs: Supervision; Project administration; Writing - review &
editing

- [1] C. Pasquini, Near infrared spectroscopy: A mature analytical technique
 with new perspectives A review, Analytica Chimica Acta 1026 (2018)
 8–36.
- [2] C. P. Bacon, Y. Mattley, R. DeFrece, Miniature spectroscopic instrumentation: Applications to biology and chemistry, Review of Scientific
 Instruments 75 (2004) 1–16.
- [3] R. V. Rossel, D. Walvoort, A. McBratney, L. J. Janik, J. Skjemstad,
 Visible, near infrared, mid infrared or combined diffuse reflectance spectroscopy for simultaneous assessment of various soil properties, Geoderma 131 (2006) 59–75.
- [4] N. Prieto, R. Roehe, P. Lavin, G. Batten, S. Andres, Application of near
 infrared reflectance spectroscopy to predict meat and meat products
 quality: A review, Meat Science 83 (2009) 175–186.
- [5] Y. Roggo, P. Chalus, L. Maurer, C. Lema-Martinez, A. Edmond,
 N. Jent, A review of near infrared spectroscopy and chemometrics in

- ⁴⁵³ pharmaceutical technologies, Journal of pharmaceutical and biomedical
 ⁴⁵⁴ analysis 44 (2007) 683–700.
- [6] V. R. Kondepati, H. M. Heise, J. Backhaus, Recent applications of nearinfrared spectroscopy in cancer diagnosis and therapy, Analytical and
 Bioanalytical Chemistry 390 (2008) 125–139.
- [7] J. H. Kalivas, Two data sets of near infrared spectra, Chemometrics
 and Intelligent Laboratory Systems 37 (1997) 255–259.
- [8] D. A. Burns, E. W. Ciurczak, Handbook of near-infrared analysis, CRC
 press, 2007.
- [9] ISO 12099:2017, Animal feeding stuffs, cereals and milled cereal products Guidelines for the application of near infrared spectrometry,
 Standard, International Organization for Standardization, Geneva, CH,
 2017.
- [10] P. Williams, P. Dardenne, P. Flinn, Tutorial: Items to be included in a
 report on a near infrared spectroscopy project, Journal of Near Infrared
 Spectroscopy 25 (2017) 85–90.
- [11] M. Larrechi, M. Callao, Strategy for introducing nir spectroscopy and
 multivariate calibration techniques in industry, TrAC Trends in Analytical Chemistry 22 (2003) 634–640.
- ⁴⁷² [12] Å. Rinnan, F. van den Berg, S. B. Engelsen, Review of the most common
 ⁴⁷³ pre-processing techniques for near-infrared spectra, TrAC Trends in
 ⁴⁷⁴ Analytical Chemistry 28 (2009) 1201–1222.

- [13] Z. Xiaobo, Z. Jiewen, M. J. Povey, M. Holmes, M. Hanpin, Variables
 selection methods in near-infrared spectroscopy, Analytica chimica acta
 667 (2010) 14–32.
- [14] R. M. Balabin, S. V. Smirnov, Variable selection in near-infrared spectroscopy: benchmarking of feature selection methods on biodiesel data,
 Analytica chimica acta 692 (2011) 63–72.
- [15] A. Candolfi, R. De Maesschalck, D. Jouan-Rimbaud, P. A. Hailey, D. L.
 Massart, The influence of data pre-processing in the pattern recognition of excipients near-infrared spectra, Journal of Pharmaceutical and Biomedical Analysis 21 (1999) 115–132.
- [16] L. J. Chen, L. Xing, L. J. Han, Influence of Data Preprocessing on the
 Quantitative Determination of Nutrient Content in Poultry Manure by
 Near Infrared Spectroscopy, Journal of Environment Quality 39 (2010)
 1841.
- [17] L. Xu, Y. P. Zhou, L. J. Tang, H. L. Wu, J. H. Jiang, G. L. Shen,
 R. Q. Yu, Ensemble preprocessing of near-infrared (NIR) spectra for
 multivariate calibration, Analytica Chimica Acta 616 (2008) 138–143.
- [18] L.-L. Wang, Y.-W. Lin, X.-F. Wang, N. Xiao, Y.-D. Xu, H.-D. Li, Q.-S.
 Xu, A selective review and comparison for interval variable selection
 in spectroscopic modeling, Chemometrics and Intelligent Laboratory
 Systems 172 (2018) 229–240.
- ⁴⁹⁶ [19] A. Mouazen, B. Kuang, J. De Baerdemaeker, H. Ramon, Comparison
 ⁴⁹⁷ among principal component, partial least squares and back propagation

neural network analyses for accuracy of measurement of selected soil
properties with visible and near infrared spectroscopy, Geoderma 158
(2010) 23–31.

- [20] R. M. Balabin, R. Z. Safieva, E. I. Lomakina, Comparison of linear and
 nonlinear calibration models based on near infrared (nir) spectroscopy
 data for gasoline properties prediction, Chemometrics and intelligent
 laboratory systems 88 (2007) 183–188.
- [21] R. M. Balabin, R. Z. Safieva, E. I. Lomakina, Gasoline classification us ing near infrared (NIR) spectroscopy data: Comparison of multivariate
 techniques, Analytica Chimica Acta 671 (2010) 27–35.
- ⁵⁰⁸ [22] J. Engel, J. Gerretzen, E. Szymańska, J. J. Jansen, G. Downey,
 L. Blanchet, L. M. Buydens, Breaking with trends in pre-processing?,
 ⁵¹⁰ TrAC Trends in Analytical Chemistry 50 (2013) 96–106.
- [23] J. Gerretzen, E. Szymańska, J. J. Jansen, J. Bart, H. J. Van Manen,
 E. R. Van Den Heuvel, L. M. Buydens, Simple and Effective Way for
 Data Preprocessing Selection Based on Design of Experiments, Analytical Chemistry 87 (2015) 12096–12103.
- ⁵¹⁵ [24] J. Gerretzen, E. Szymańska, J. Bart, A. N. Davies, H. J. van Manen,
 ⁵¹⁶ E. R. van den Heuvel, J. J. Jansen, L. M. Buydens, Boosting model
 ⁵¹⁷ performance and interpretation by entangling preprocessing selection
 ⁵¹⁸ and variable selection, Analytica Chimica Acta 938 (2016) 44–52.
- 519 [25] T. Bocklitz, A. Walter, K. Hartmann, P. Rösch, J. Popp, How to pre-

- process Raman spectra for reliable and stable models?, Analytica Chimica Acta 704 (2011) 47–56.
- ⁵²² [26] F. Pedregosa, G. Varoquaux, A. Gramfort, V. Michel, B. Thirion,
 ⁵²³ O. Grisel, M. Blondel, P. Prettenhofer, R. Weiss, V. Dubourg, et al.,
 ⁵²⁴ Scikit-learn: Machine learning in python, Journal of machine learning
 ⁵²⁵ research 12 (2011) 2825–2830.
- ⁵²⁶ [27] E. Jones, T. Oliphant, P. Peterson, et al., Scipy: Open source scientific ⁵²⁷ tools for python (2001).
- [28] R. J. Barnes, M. S. Dhanoa, S. J. Lister, Standard normal variate transformation and de-trending of near-infrared diffuse reflectance spectra,
 Applied Spectroscopy 43 (1989) 772–777.
- [29] H. Martens, S. Jensen, P. Geladi, Multivariate linearity transformation
 for near-infrared reflectance spectrometry, in: Proceedings of the Nordic
 symposium on applied statistics, Stokkand Forlag Publishers Stavanger,
 Norway, pp. 205–234.
- [30] Q. Guo, W. Wu, D. L. Massart, The robust normal variate transform
 for pattern recognition with near-infrared data, Analytica Chimica Acta
 382 (1999) 87–103.
- [31] Y. Bi, K. Yuan, W. Xiao, J. Wu, C. Shi, J. Xia, G. Chu, G. Zhang,
 G. Zhou, A local pre-processing method for near-infrared spectra, combined with spectral segmentation and standard normal variate transformation, Analytica Chimica Acta 909 (2016) 30–40.

- [32] H. Martens, E. Stark, Extended multiplicative signal correction and
 spectral interference subtraction: new preprocessing methods for near
 infrared spectroscopy, Journal of pharmaceutical and biomedical analysis 9 (1991) 625-635.
- [33] N. K. Afseth, A. Kohler, Extended multiplicative signal correction in
 vibrational spectroscopy, a tutorial, Chemometrics and Intelligent Laboratory Systems 117 (2012) 92–99.
- [34] A. Savitzky, M. J. Golay, Smoothing and differentiation of data by simplified least squares procedures., Analytical chemistry 36 (1964) 1627–
 1639.
- [35] L. A. Sordillo, Y. Pu, S. Pratavieira, Y. Budansky, R. R. Alfano, Deep
 optical imaging of tissue using the second and third near-infrared spectral windows, Journal of Biomedical Optics 19 (2014) 056004.
- ⁵⁵⁵ [36] F. Kosmowski, T. Worku, Evaluation of a miniaturized nir spectrometer
 ⁵⁵⁶ for cultivar identification: The case of barley, chickpea and sorghum in
 ⁵⁵⁷ ethiopia, PloS one 13 (2018) e0193620.
- [37] J. K. Sarin, M. Amissah, H. Brommer, D. Argüelles, J. Töyräs, I. O.
 Afara, Near infrared spectroscopic mapping of functional properties of
 equine articular cartilage, Annals of biomedical engineering 44 (2016)
 3335–3345.
- [38] M. Prakash, J. K. Sarin, L. Rieppo, I. O. Afara, J. Töyräs, Optimal
 regression method for near-infrared spectroscopic evaluation of articular
 cartilage, Applied spectroscopy 71 (2017) 2253–2262.

- ⁵⁶⁵ [39] J. K. Sarin, J. Torniainen, M. Prakash, L. Rieppo, I. O. Afara, J. Töyräs,
 ⁵⁶⁶ Dataset on equine cartilage near infrared spectra, composition, and func⁵⁶⁷ tional properties, Scientific Data 6 (2019) 164.
- ⁵⁶⁸ [40] A. Stevens, L. Ramirez-Lopez, An introduction to the prospectr package ⁵⁶⁹ (2013) 1–22.
- [41] R. A. Viscarra Rossel, ParLeS: Software for chemometric analysis of
 spectroscopic data, Chemometrics and Intelligent Laboratory Systems
 90 (2008) 72–83.
- ⁵⁷³ [42] S. Wold, H. Antti, F. Lindgren, J. Öhman, Orthogonal signal correc⁵⁷⁴ tion of near-infrared spectra, Chemometrics and Intelligent Laboratory
 ⁵⁷⁵ Systems 44 (1998) 175–185.
- ⁵⁷⁶ [43] M. Feurer, A. Klein, K. Eggensperger, J. Springenberg, M. Blum,
 ⁵⁷⁷ F. Hutter, Efficient and robust automated machine learning, in: Ad⁵⁷⁸ vances in Neural Information Processing Systems, pp. 2962–2970.
- [44] R. S. Olson, J. H. Moore, Tpot: A tree-based pipeline optimization tool
 for automating machine learning, in: Workshop on Automatic Machine
 Learning, pp. 66–74.

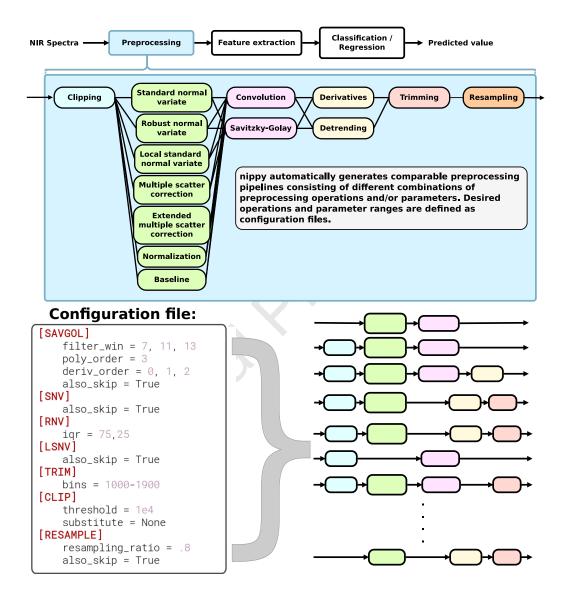


Figure 1: General operation principle of nippy. Several preprocessing options and parameter values can be combined into multiple competing preprocessing strategies (called pipelines). Comparing the effect of different preprocessors on the prediction performance of the NIRS model will yield the most optimal solution for a given application.

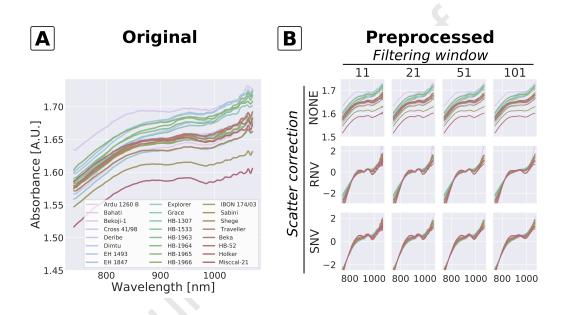


Figure 2: A: Overview of the Etiophian barley NIRS data set. Individual lines correspond to the averaged unprocessed spectra of the 24 different barley variants. B: The effect of different parameter combinations of scatter correction and smoothing to the per-class average spectra of the dataset. Rows correspond to different scatter correction treatment while columns represent different filter window lengths used in Savitzky-Golay filtering (3rd polynomial order, no derivation).

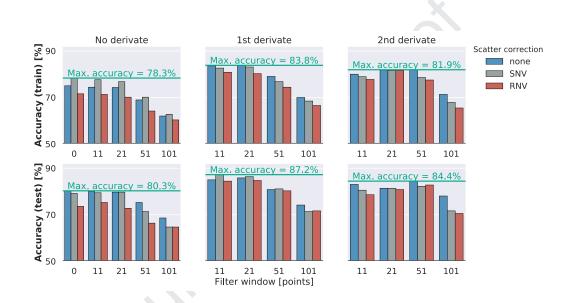


Figure 3: Overall training and testing accuracy with SVM classification for barley cultivars of example 1. Training accuracy was derived from cross-validation while testing accuracy represents the performance of an independent hold-out set. Results have been divided between filtering window length (the effect of omitting filtering was also investigated), scatter correction method, and derivative.

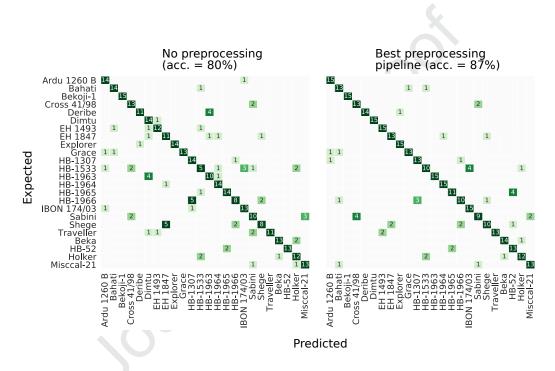


Figure 4: A comparison of confusion matrices between the un-preprocessed data and the data preprocessed with the optimal preprocessing combination of the barley cultivar classification example.

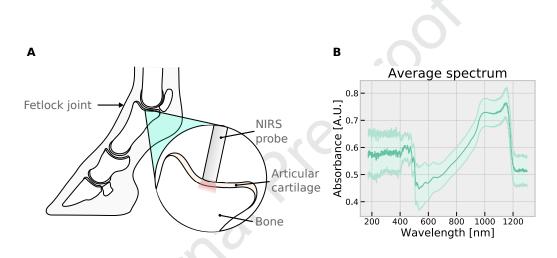


Figure 5: A: NIRS spectra was measured from the cartilage surface of the equine fetlock joint. In total, five joints were measured from 44 different areas of interest (AIs). Each AI consisted between 6–25 measurement points (depending on the joint geometry and cartilage condition) resulting in 861 measurement points. Local instantaneous modulus was determined for each measurement site. B: Average of all the collected NIR spectra. Shaded regions represent the standard deviation.

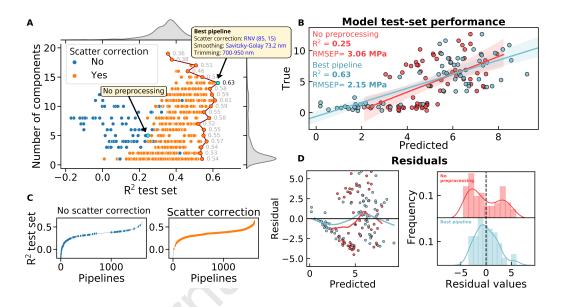
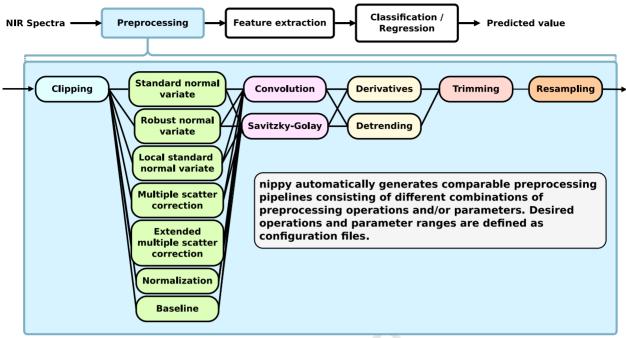
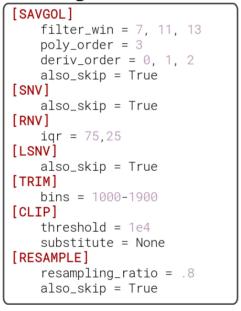
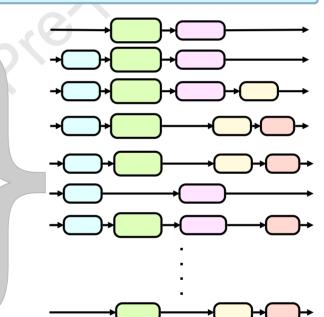


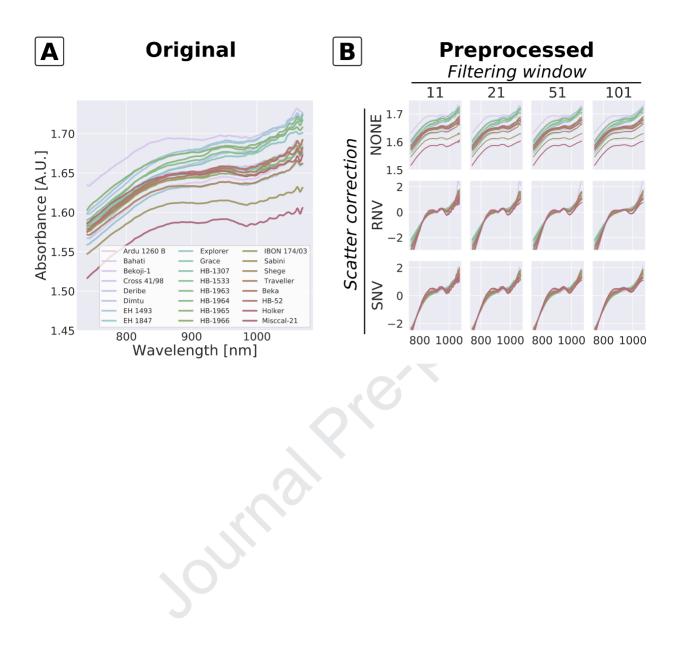
Figure 6: A: All 1618 models generated to predict the instantaneous modulus of articular cartilage presented in terms of model complexity (i.e., the number of latent variables) and test set performance. Color of individual models indicates whether the data was treated with scatter correction. Margin plots indicate the distribution of models in terms of complexity and performance. B: True vs predicted values for the test set between no preprocessing and the best performing pipeline. C: All preprocessing pipelines (scatter correction vs no scatter correction) sorted according to the \mathbb{R}^2 values of the test set. D: Residual plots and the distributions of residuals between no preprocessing and the best performing pipeline.

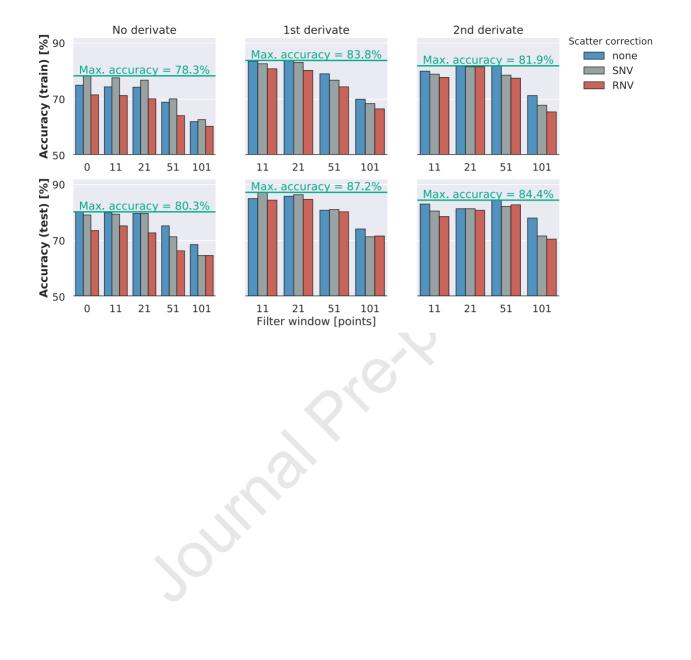


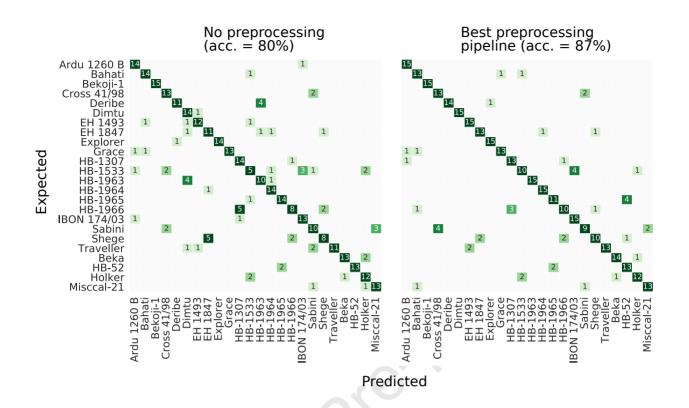
Configuration file:



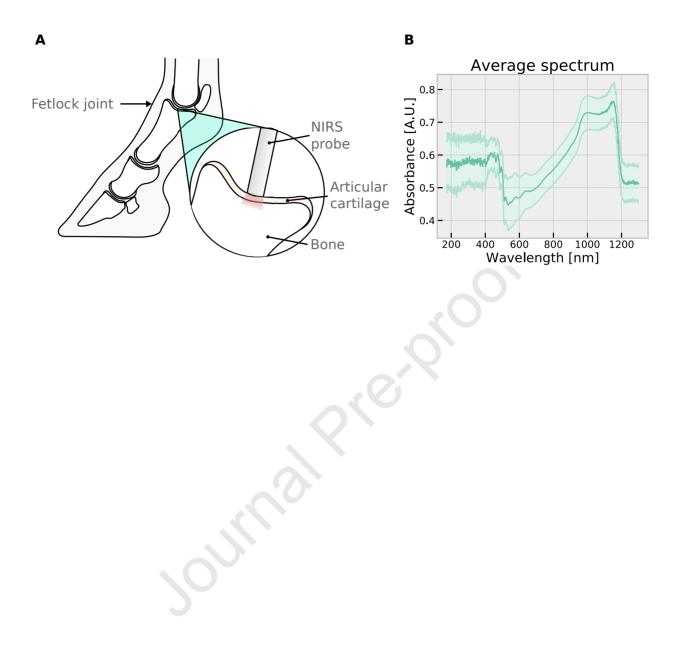


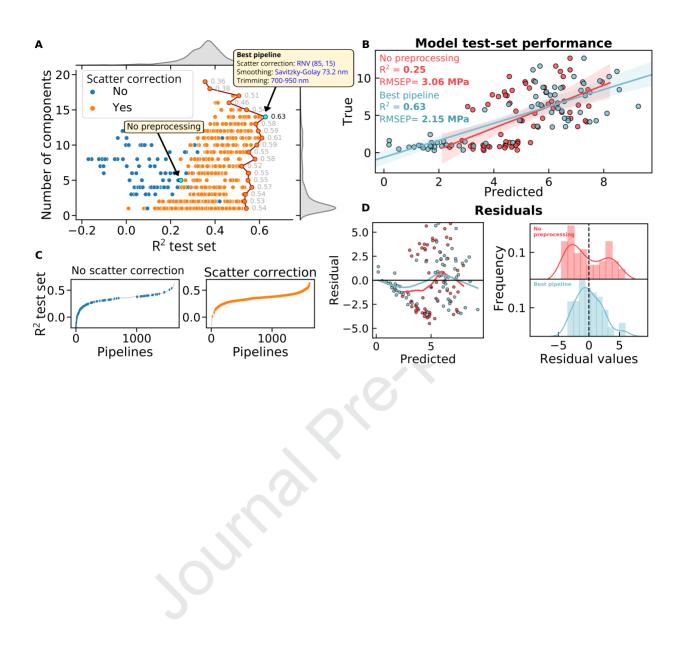






Jonus





Highlights

- Spectral preprocessing affects the performance of chemometric ٠ models
- Selection of best preprocessing strategy depends heavily on the • intended application
- nippy module can rapidly explore different preprocessing • combinations
- Semi-automatic preprocessing tuning for spectroscopic models

Declaration of interests

X The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

of all the authors; On beha JARI TORNIAINEN